

Hydrophilic interactions; their Origin, their Magnitude and their Relevance to Protein Folding

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Abstract

A simple model is constructed in which the phenomenon of hydrophilic $(H\phi I)$ interactions may be studied exactly. This model reveals the *origin* of the strong $H\phi I$ interaction, as well as providing an approximate method of estimating its order of magnitude. The results are in agreementwith previous theoretical calculations, as well as with experimental data, and simulated calculations. The source of the non-additivity of the $H\phi I$ interactions among three and four $H\phi I$ interactions is examined. A few comments on the relevance of the $H\phi I$ interactions to the process of protein folding are discussed.

Key words: Hydrophobic Hydrophilic Interactions. Hydrogen Bonds. Protein Folding.



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1. Introduction

In the late 1980s and early 1990s, a systematic study of all possible solvent-induced effects on protein folding was undertaken.¹⁻¹⁰ One of the most unexpected discovery was that hydrophilic ($H\phi I$) interactions, as defined in the next section, are much stronger than the corresponding hydrophobic ($H\phi O$) interactions.

These findings have a profound effect on our understanding of some fundamental biochemical processes such as protein folding, protein-protein association and molecular recognition.¹¹⁻¹² Yet, in spite of the overwhelming evidence for these findings (from theoretical, experimental and simulations), they were largely ignored. If you open any modern textbook of biochemistry, biophysics or molecular biology, you will find statements about the "major importance of the $H\phi O$ effect," and on the "insignificant contribution of hydrogen bonding" in protein folding.¹²

I believe that there are two main reasons for the continual lingering of the myth about the importance of the $H\phi O$ in protein folding.

The first is that most people in the field, including myself, have spent many years in studying, teaching and applying the $H\phi 0$ effect, and it is very hard to deviate from the accepted paradigm regarding the dominance of the $H\phi 0$ effect, even after strong evidence favoring the new paradigm involving $H\phi I$ effects became available.

The second reason is that both $H\phi O$ and $H\phi I$ effects are difficult to understand. As solvent-induced interaction the understanding of these effects depend on the knowledge of the notoriously difficult statistical mechanical theory of liquid water.

In 1959, Kauzmann¹³ had proposed a simple, intuitive and very appealing, some even claim convincing¹² model for the $H\phi 0$ effect. This was the Gibbs energy change for the process of transferring a non-polar solute from water into an organic liquid. It was known that the Gibbs energy change in this transfer process is large and negative. Kauzmann realized the analogy between this transfer process and the process of transferring a $H\phi 0$ group of a protein from being exposed to water into the interior of a protein, Figure 1. If each of these $H\phi 0$ groups of a protein contributes about -2 to -3 kcal/mol then one can expect a large negative change to the Gibbs energy of the process of protein folding.

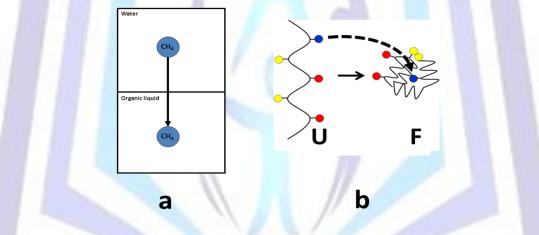


Figure 1. Kauzmann's model: a) The process of transferring a non-polar solute from water into an organic liquid. b) The transferring of a non-polar group (blue) into the interior of the protein.

Kauzmann's model had captured the imagination and the conviction of most scientists.¹⁴ No doubt this is the reason that most textbooks claim that the $H\phi O$ effect is the most important one in the process of protein folding.

In 1990, I have shown that Kauzmann's model was not adequate to the problem of protein folding.² It was also shown^{1,5} that $H\phi I$ effects might be more important to protein folding. Unfortunately, the argument in favor of the $H\phi I$ effects were not easy to grasp and to accept. There has also been much confusion in identifying the $H\phi I$ effects with direct intra-molecular hydrogen bonding (HB).

The purpose of this article is to first clarify the difference between various solvent induced effects involving $H\phi I$ groups, second, to reveal the origin of the strong $H\phi I$ effects, and to provide an estimate of the order of magnitude of these effects. Finally, we shall comment on the relevance of $H\phi I$ effects to protein folding.

To achieve this goal a simple and solvable model has been designed. This is described in section 3. It should be stressed that this is not a model for aqueous solution. The purpose of the model is to clarify the *origin* of the $H\phi I$ interactions. In section 4, we discuss the $H\phi I$ interactions in real aqueous solutions. We show that the essence of the argument for the existence of strong $H\phi I$ in real systems is almost the same as in the simple model. Some concluding remarks are discussed in section 5.



2. Definitions and motivation for the constructing of a model for $H\phi I$

interactions

We consider a system of N water molecules at some temperature T and pressure P, and two solutes 1 and 2 at some fixed positions, and orientations in the solvent. The process of interest is to bring these two solutes from infinite separation $R = \infty$ to some close distance R, Figure 2.

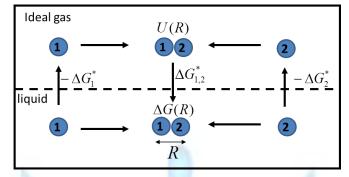


Figure 2. The process of bringing two solutes from fixed position at infinite separation to the final distance R. If the process is carried out both in an ideal gas phase and in the liquid phase, one gets the relationship between the solvent induced part of the Gibbs energy change and the solvation Gibbs energies of all the solutes involved.

The Gibbs energy change for this process is the same as the potential of mean force (PMF) between the two solutes^{1,12}. This can be written as

$$\Delta G(R) = W(R) = U(R) + \delta G$$

$$= \Delta U(R) + \Delta G_{1,2}^* - \Delta G_1^* - \Delta G_2^*$$
(2.1)

where U(R) is the pair potential between the two solutes in vacuum. Usually, we also need to specify the relative orientations of the two solutes but this will be done for each of the $H\phi I$ effects separately. δG is referred to as the solvent-induced contribution to the Gibbs energy change. The latter can always be expressed in terms of solvation Gibbs energies of all the species involved in the process. The second equality can be derived either theoretically¹² or simply from cyclic process in Figure 2.

When the solutes are hydrophobic $(H\phi 0)$, say methane, ethane and the like, the quantity δG may be referred to as $H\phi 0$ interaction. This effect has been discussed in great detail in references 11 and 12 and will not be discussed here. When the solutes are hydrophilic $(H\phi I)$, such as methanol, ethanol, etc., we refer to δG as the $H\phi I$ interaction. We also apply the quantity δG for two $H\phi I$ groups, say hydroxyl, carbonyl or amine, attached to some backbone molecule. This is important in the application of the $H\phi I$ to protein folding.

We shall next specify several $H\phi I$ effects, some of these will be further studied within the model described in the next section.

A. Formation of direct hydrogen bond(HB) between the two solutes

The relevant process is depicted in Figure 3.

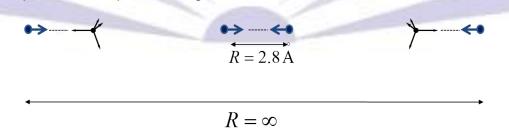


Figure 3. The process of forming a direct HB between two $H\phi I$ groups in the water.

Two solutes are brought from infinite separation to a distance of about $R \sim 2.8$ Å, and with the correct orientations such that an HB is formed between the two solutes. The Gibbs energy change in this process is

$$\Delta G^{HB} = U(R) + \delta G(R)$$
$$= \epsilon_{HB} + \Delta G_{1,2}^* - \Delta G_1^* - \Delta G_2^*$$
$$\approx \epsilon_{HB} - \Delta G^* (one \ arm \ of \ 1) - \Delta G^* (one \ arm \ of \ 2) \qquad (2.2)$$



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In this process the direct interaction is approximately equal to a HB energy, denoted ϵ_{HB} . The solvent induced part is essentially equal to the loss of the solvation Gibbs energy of one "arm" on solute 1, and one "arm" on solute 2. This approximation is equivalent to viewing the $H\phi I$ solute as consisting of a few "arms" along which a hydrogen bond may be formed. For instance, amine group NH has one arm, carbonyl group, C = 0 has two arms, hydroxyl group, OH has three arms and a water molecule has four arms. In writing the approximation (2.2) it is assumed that only the solvation of the arms involved in the formation of HB should be taken into account. It is also assumed that the solvations of different arms on a single solute are independent.¹⁵ Other parts of the molecules, the environment of which are unchanged will not contribute to the Gibbs energy change of the process.¹ Of course, in reality one needs to distinguish between a donor and an acceptor arms, but for the purpose of this article we ignore this distinction.

The solvation Gibbs energy of one arm was estimated based on a simple model for water, as well as from experimental data to be about -2.25 kcal/mol.^{11,12} Taking ϵ_{HB} to be of the order of -6 to -6.5 kcal/mol, we get for ΔG in (2.2)

$$\Delta G^{HB} \cong -6.5 + 2 \times 25 \approx -2 \ kcal/mol \tag{2.3}$$

This is significantly different from what is inferred from the so-called HB inventory argument which was discussed in great detail in reference 11, 12 and 16. It should be noted that most people when discussing $H\phi I$ effects in protein folding are actually referring to the process of formation of an intra-molecular HB by $H\phi I$ groups. Thus, the HB inventory argument has not only underestimated the contribution of intra-molecular HBing, but also has led to the dismissal of other, possibly more important $H\phi I$ effects as discussed below.

B. $H\phi I$ interaction between two $H\phi I$ groups at a distance of about 4.5Å

This is perhaps the most important $H\phi I$ effect in both protein folding and protein-protein association.^{11,12} Here, we consider the process of bringing two solutes from infinite separation to a distance $R \approx 4.5 \text{\AA}$, within the solvent. We also require that the orientations of these solutes is such that one arm of solute 1 and one arm of solute 2 can form HBs with one water molecule, Figure 4.

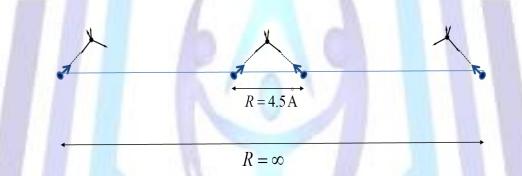


Figure 4. The process, referred to as pairwise $H\phi I$ interaction. Two arms of either two solutes or one of two groups on protein are brought from infinite separation to a distance R = 4.5Å, and an appropriate orientation so that they can form HBs with one water molecule.

The Gibbs energy change for this process is

$$\Delta G_2^{H\phi I} \left(R \approx 4.5 \text{\AA} \right) = \Delta U + \delta G_2^{H\phi I}$$

 $\approx \Delta G_{1,2}^*(of \text{ two arms at the final configuration}) - \Delta G_1^*(one \text{ arm of solute 1}) - \Delta G_2^*(one \text{ arm of solute 2})$ (2.4)

In this process we assume that the *direct* interaction between the two $H\phi I$ groups is relatively small. As before, the solvent-induced part of ΔG is due only to the solvation of those parts of the solute which are changed in the process, specifically, the two arms on the two solutes. As in the process depicted in Figure 3, we assume that the types of arms (donor or acceptor) match when a HB is formed.

If we look superficially at the process as depicted in Figure 4, we see in both the initial and the final states, that the arm of solute 1 can form a HB with water molecule, and the arm of solute 2 can form a HB with water molecule. It is easy to conclude that the solvation of the two solutes will not change in the process, hence there is no reason to expect any significant solvent-induced effect in this process. We shall see in the next section that this conclusion is false. It arises from confusing a HB *energy* with *solvation Gibbs energy* of a HBing arm. This is the same error committed in the HB-inventory argument,¹² and the main reason that has hindered understanding this $H\phi I$ effect.

C. $H\phi I$ interaction between three $H\phi I$ groups

As an extension of the pairwise $H\phi I$ interaction discussed above considers the following process. We start with three $H\phi I$ solutes (or groups on a protein) initially at infinite separation from each other, to a final configuration as shown in Figure 5.



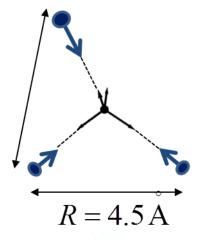


Figure 5. $H\phi I$ process between three arms belonging to either three solutes or to three $H\phi I$ groups on a protein. Only the final configuration is shown.

At this configuration the three arms of the three $H\phi I$ solutes can form three HBs with a water molecule. The Gibbs energy change for this process is

$$\Delta G_3^{H\phi l} = \Delta U(1,2,3) + \delta G_3^{H\phi l} \tag{6}$$

Again, we can assume that the *direct* interaction between the three solutes is relatively small and the main contribution to the Gibbs energy change is the indirect, solvent-induced part denoted $\delta G_3^{H\phi l}$.

2.5)

A superficial examination of this process could lead to the erroneous conclusion that $\delta G_3^{H\phi I}$ should be negligible. The argument; three arms of the solutes form three HBs with water molecules in both the initial and the final state. Therefore, one would not expect a large contribution from this effect. As before the fallacy of this conclusion is confusing a HB *energy* with *solvation Gibbs energy* of a HBing arm of a $H\phi I$ solute (or group). We shall see in the next section the origin of this confusion, and also find that $\delta G_3^{H\phi I}$ is even larger than $\delta G_2^{H\phi I}$. We shall also see that $\delta G_3^{H\phi I}$ is not pairwise additive.

D. $H\phi I$ interaction between four $H\phi I$ groups

The final process that we shall study in the next section is the $H\phi I$ interaction between four $H\phi I$ solutes or groups. We consider the process of bringing four $H\phi I$ solutes from infinite separation to the final configuration as depicted in Figure 6.

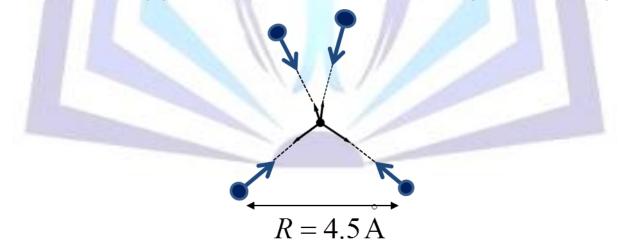


Figure 6. $H\phi I$ process between four arms belonging to either four solutes or to three $H\phi I$ groups on a protein. Only the final configuration is shown.

The four $H\phi I$ solutes in the final configuration can form four HBs with a water molecule. The corresponding Gibbs energy change is

$$\Delta G_4^{H\phi I} = \Delta U(1,2,3,4) + \delta G_4^{H\phi I}$$
(2.6)

We shall study this $H\phi I$ interaction in the next section within a simple model system. We shall see that this effect is larger than the $H\phi I$ interaction between three $H\phi I$ groups. We shall also see that this $H\phi I$ interaction is not pairwise additive.



E. Other stronger and longer range $H\phi I$ interactions

There is a host of other solvent induced interactions, some are stronger than the ones discussed above, and some have longer range. We shall not discuss these in this article. They were discussed in detail in reference 12.

3. A "minimal" model showing the origin of the $H\phi I$ interactions

The model described below was specifically designed so that it is simple enough to be exactly solvable, yet contains enough features that enable us to study the *molecular origin* of the $H\phi I$ effects, to estimate the order of magnitude of these effects, and to examine the source of the non-additivity of the $H\phi I$ effects. It should be emphasized, however that we do not consider this model to be reliable for estimating the $H\phi I$ effects in real aqueous solutions. It is used here for demonstrating the *origin* of the various effects. The estimated values of the $H\phi I$ effects will be done in the next section (see also reference 12).

As we have seen in the previous section all of the $H\phi I$ interactions may be expressed as differences in solvation Gibbs energies of solutes at various configurations. Each solvation Gibbs energy of a solute *s* has the form^{11,12}

$$\Delta G_s^* = -k_B T ln \langle exp[-\beta B_s] \rangle_0 \tag{3.1}$$

Here, k_B is the Boltzmann constant, T the absolute temperature. $\beta = (k_B T)^{-1}$. B_s is the binding energy, i.e. the total interaction energy of the solute *s* with all solvent molecules at a specific configuration $X^N = X_1, \dots, X_N$, i.e.

$$B_s(\boldsymbol{X}_s) = \sum_{i=1}^N U(\boldsymbol{X}_s, \boldsymbol{X}_i)$$
(3.2)

where $U(X_s, X_i)$ is the solute-solvent pair potential.

The average $\langle \rangle_0$ in (3.1) is over all possible configurations of the solvent molecules with the probability distribution of water molecules in the absence of the solute *s*.

$$\langle exp[-\beta B_s] \rangle_0 = \int \cdots \int exp[-\beta B_s] P_0(\mathbf{X}^N) d\mathbf{X}^N$$
(3.3)

where

$$P_0(\mathbf{X}^N) = \frac{\exp\left[-\beta U_N(\mathbf{X}^N)\right]}{\int \cdots \int \exp\left[-\beta U_N(\mathbf{X}^N)\right] d\mathbf{X}^N}$$
(3.4)

In (3.3) we have taken the average in the (T, V, N) ensemble, whereas in (3.1) one should take the average in the T, P, N ensemble. However, in actual aqueous systems we can neglect the difference between the Gibbs and the Helmholtz energy of the solvation.¹¹

As we can see from (3.1) the solvation Gibbs energy depends on the solute-solvent interaction through B_s , and the solvent-solvent interaction through the distribution function $P_0(X^N)$ in (3.4). For the study of the $H\phi I$ interactions we need to examine the *difference* between solvation Gibbs energies. In such difference the more important factor is the solute-solvent interaction. Therefore, in the model described below we take into account only solute-solvent interactions and neglect solvent-solvent interaction. The latter assumption renders the solvability of the model.

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Figure 7.A 2D system of M cells, and N water-like molecules.

We consider a lattice system of M cells and N water-like molecules (N<M). Figure 7 shows a 2D depiction of the system. Each cell can be occupied only by one water molecule at a fixed point, i.e. it lacks translational degrees of freedom. Each water molecule has four arms along which it can form HB with an arm of a solute molecule, but we neglect HBing between water molecules. We further assume that each water molecule can be in one of *n* different orientations. For example, Figure 8 shows the case of *n*=10 and the configurations are described by the angle α_i

$$\alpha_i = i \frac{\pi}{40} \tag{3.5}$$



 \neq 0 corresponds to the configuration on the lhs of Figure 8, and \neq 10 corresponds to a rotation by an angle of $\pi/4$, which brings the molecule to the equivalent configuration of \neq 0.

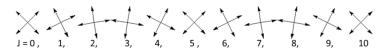


Figure 8. All possible orientations of the "water" molecule with n=10. i=0 corresponds to the left hand configuration, i=10 is identical to i=0.

The canonical partition function (PF) of the pure solvent is thus

 $Q_0 = {M \choose N} q^N$, where q=n (3.6)

We now place a solute at a fixed cell. The solute has one arm along which it can form a HB with another solute or with one water molecule being in the correct orientation.

For a system of N water molecules and one solute at a fixed cell and having a fixed orientation, the partition function is

$$Q_1 = \sum_i exp[-\beta\epsilon_i] \tag{3.7}$$

In this system there are only two energy levels; either the solute forms a HB with a water molecule, or not. We assume that the water molecule can form a HB with the solute molecule only when it is in one out of the *n* orientations, say \models 0 in Figure 8. In this case, the PF has the form

$$Q_1 = {\binom{M-2}{N-1}} n^{N-1} exp[-\beta \epsilon_{HB}] + {\binom{M-2}{N-1}} n^{N-1} (n-1) + {\binom{M-2}{N}} n^N$$
(3.8)

The first terms on the rhs of (3.8) corresponds to all configurations for which a HB is formed between the solute and a water molecule. The second term corresponds to all configurations for which a water molecule is in the cell next to the arm of the solute but it does not form HB. The third term is the total number of configurations for which there is no water molecule at the site next to the arm of the solute.

The Gibbs energy change for placing a solute at a fixed cell in this system is thus

$$\Delta G_s^*(at \ a \ fixed \ cell) = -k_B T ln \frac{Q_1}{Q_1}$$
(3.9)

For very large N and M we get from (3.9)

 $\Delta G_s^*(at \ a \ fixed \ cell) = -k_B T ln(1-\theta)$

 $-k_B T ln \left[exp[-\beta \varepsilon_{HB}] \frac{\theta}{n} + 1 - \frac{\theta}{n} \right]$ (3.10)

where $\theta = N/M$ is the fraction of cells occupied by water molecules. This is also the probability of finding a water molecule at a specific cell. The first term on the rhs of (3.10) is the Gibbs energy change to form a "cavity" in the system, which in our case is an empty cell. This is also the limit of ΔG_s^* in (3.10) for the case $\varepsilon_{HB} = 0$, hence

$$\Delta G_s^*(at \ a \ fixed \ cell, and \ \varepsilon_{HB} = 0) = -k_B T ln(1-\theta)$$
(3.11)

In a real solvent this is the so-called "cavity work".^{11,12} Here, it is the work required to find a cell empty. This is zero when $\theta \rightarrow 0$, but it is infinite when $\theta \rightarrow 1$, i.e. all cells are occupied. Note also that this quantity is essentially entropic, and it does not depend on the interactions in the system.

For the next analysis we shall not need the "cavity work" term. This will always cancel out in the process of $H\phi I$ interactions between $H\phi I$ groups on a protein. We always take into account the conditional solvation Gibbs energy of the group^{11,12}, see also section 4.

We now examine the Gibbs energy change for the processes discussed in section 2. To get an order of magnitude for the different terms involved we rewrite the solvation Gibbs energy of one arm of a solute as

$$\Delta G_1^*(one \ arm) = -k_B T ln\{exp[-\beta \varepsilon_{HB}]P_{HB} + (1 - P_{HB})\}$$
(3.12)

where $P_{HB} = \theta/n$ is the probability of finding a water molecule at the right cell next to the solute's arm $\left(\theta = \frac{N}{M}\right)$, and that the water molecule is in the right configuration to form a HB, $\left(\frac{1}{n}\right)$. To get a feel for an order of magnitude of the various terms in (3.12) we take $R = 1.987 cal/mol \ K,T = 298K$ and $\varepsilon_{HB} \approx -6.5 \ kcal/mol$. We also know the experimental, as well as the theoretical value of the (conditional) Gibbs energy of solvation of one arm which is about $-2.25 \ kcal/mol$.^{11,12} Therefore, from the equality

$$-RTln\{exp[-\beta \varepsilon_{HB}]P_{HB} + (1 - P_{HB})\} = -2.25 \ kcal/mol$$
(3.13)



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we get, for $\varepsilon_{HB} = -6.5kcal/mol$

$$P_{HB} \approx 0.0007$$

and for $\varepsilon_{HB} = -6 \ kcal/mol$

$$P_{HB} \approx 0.0017$$

For the process of $H\phi I$ interactions between two arms as depicted in Figure 9, we have

$$\delta G_{2}^{H\phi I} = -k_{B}T ln \left\{ \frac{exp \left[-\beta 2 \varepsilon_{HB} \right] P_{HB} + (1 - P_{HB}) \right]}{[exp \left[-\beta \varepsilon_{HB} \right] P_{HB} + (1 - P_{HB})]^{2}} \right\}$$
(3.14)

Figure 9. The process of pairwise $H\phi I$ interaction between two arms of two solutes in the model system.

First we examine two limiting cases:

1. When $P_{HB} \rightarrow 0$. This can occur either when the density of water is too low $(\theta \rightarrow 0)$, or that *n* is very large $(\frac{1}{n} \rightarrow 0)$. In this limit, as expected, there is no solvent-induced effect, i.e. $\delta G_2^{H\phi I} \rightarrow 0$

2. When $P_{HB} \rightarrow 1$. This can occur when nearly all the cells are occupied by water molecules ($\theta \approx 1$), and that there is only one configuration (n = 1). In this limit we also have

$$\delta G_2^{H\phi I} = -k_B T ln \left[\frac{exp \left[-\beta 2\varepsilon_{HB} \right]}{exp \left[-\beta 2\varepsilon_{HB} \right]} \right] = 0$$
(3.15)

If each arm forms a HB in both the initial and the final configuration, then we break to HBs and make two HBs and therefore the net effect is $\delta G = 0$. This is essentially the origin of the confusion that exists between the formation of a HB and the solvation by HBs.

In reality the HB energy is different from the Gibbs energy of solvation of an arm. To see the origin of the $H\phi I$ interactions we take the case of $P_{HB} \ll 1$, but $exp[-\beta \varepsilon_{HB}]P_{HB} \gg 1$, hence (3.14) reduces to

$$\delta G_2^{H\phi I} = -k_B T ln \left\{ \frac{exp \left[-\beta 2\varepsilon_{HB} \right] P_{HB}}{exp \left[-\beta 2\varepsilon_{HB} \right] P_{HB}^2} \right\} = +k_B T ln P_{HB}$$
(3.16)

Thus, for small value of P_{HB} (but not too small), we see that the main origin of the $H\phi I$ effect is that in the initial state the solvation Gibbs energies of the two solutes involve the factor $exp[-\beta 2\varepsilon_{HB}]$, and the probability P_{HB}^2 , i.e. *two* water molecules must be in the correct configuration to form HBs with the two solutes. On the other hand, in the final configuration we also have the factor $exp[-\beta 2\varepsilon_{HB}]$, but now only *one* probability factor P_{HB} .

The case discussed above reveals the main origin of the occurrence of $H\phi I$ interaction. We shall further discuss the general case in the next section. Thus, if we take the value of P_{HB} estimated from (3.12) we get for the $H\phi I$ interaction in (3.16), for $\varepsilon_{HB} = -6kcal/mol$

$$\delta G_2^{H\phi l} = k_B T ln P_{HB} \approx -3.7 k cal/mol$$
(3.17)

and for $\varepsilon_{HB} = -6.5 \ kcal/mol$

$$\delta G_2^{H\phi I} = k_B T ln P_{HB} \approx -4.3 k cal/mol$$

This result is quite significant. It shows that for a HB energy of about $\varepsilon_{HB} = -6 \ kcal/mol$, the $H\phi I$ interaction is of the order of about $-3 \ kcal/mol$. This is quite similar to the value of the $H\phi I$ interaction which was estimated from either theory or experimental data^{11,12}. For more realistic estimates see next section.



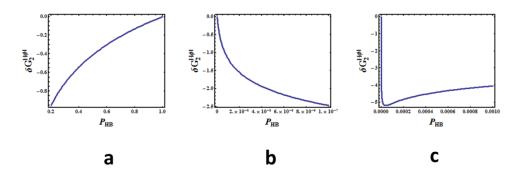


Figure 10. Values of $\delta G_2^{H\phi l}$ calculated for T = 298K and $\varepsilon_{HB} = -6kcal/mol$, as a function of the probability P_{HB} . a) For large P_{HB} , b) For very small P_{HB} , and c) For the range $0 \le P_{HB} \le 0.001$.

Figure 10 shows the values of $\delta G_2^{H\phi I}$ as a function of P_{HB} for T = 298K and $\varepsilon_{HB} = -6 \ kcal/mol$. We see that in both limits $P_{HB} \rightarrow 0$, and $P_{HB} \rightarrow 1\delta G_2^{H\phi I}$ is zero. However, for small values of P_{HB} , we can obtain strong $H\phi I$ interactions, which are of the same order of magnitude as the HB energy.

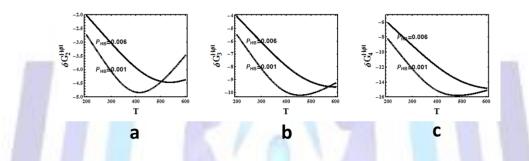


Figure 11. a) The temperature dependence of $\delta G_2^{H\phi I}$ for two values of P_{HB} . b) The same as *a*, but for $\delta G_3^{H\phi I}$. c) The same as *a*, but for $\delta G_4^{H\phi I}$.

Figure 11a shows the dependence of $\delta G_2^{H\phi I}$ on the temperature for two values of P_{HB} .

We next examine the $H\phi I$ interaction between three solutes, or three arms on three groups of a protein. As before, in the two limits $P_{HB} \rightarrow 0$, and $P_{HB} \rightarrow 1$ we have: $\delta G_3^{H\phi I} \rightarrow 0$.

However, for a small value of P_{HB} , but not too small we can write $\delta G_3^{H\phi I}$ as

$$\delta G_3^{H\phi I} = -k_B T ln \left\{ \frac{exp \left[-\beta \, 3 \varepsilon_{HB} \right] P_{HB}}{exp \left[-\beta \, 3 \varepsilon_{HB} \right] P_{HB}^3} \right\} = k_B T ln P_{HB}^2 \tag{3.18}$$

From (3.18) we see the main origin of the H_{dl} interaction between the three H_{dl} solutes, or H_{dl} groups. As in eq. (3.16) we have here three HBs in the initial and the final states. However, the probability of finding water molecules in the correct position and orientation is P_{HB} in the final state, but P_{HB}^3 in the initial state.

Figure 11b shows some values of \mathcal{X}_{3}^{HM} . Note that the values of \mathcal{X}_{3}^{HM} are about twice as large compared with \mathcal{X}_{2}^{HM} .

For the Hold interaction between four Hold groups we have approximately

$$\delta G_3^{H\phi I} = -k_B T ln \left\{ \frac{exp \left[-\beta \Im \varepsilon_{HB} \right] P_{HB}}{exp \left[-\beta \Im \varepsilon_{HB} \right] P_{HB}^3} \right\} = k_B T ln P_{HB}^3$$
(3.19)

Finally, we address the question of additivity of the H_{α} interaction. This is an important aspect of any solventinduced effect. It has been discussed in connection with the H_{α} interactions in reference 16. Here, we examine the nonadditivity of the H_{α} interaction within the simple model system.

We define the extent of non-additivity with respect to pairs of Hot groups as

$$\Delta G^3_{NON - ADD} = \delta G^{H \phi I}_3 - 3 \delta G^{H \phi I}_2$$



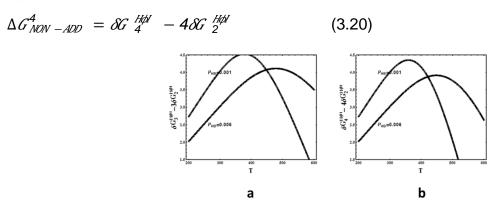


Figure 12. The extent of non additivity of a) δG_{3}^{Hold} and b) δG_{4}^{Hold} .

Figure 12 shows the values of the extent of non-additivity for three and four $H_{\mathcal{D}}$ groups. As can be seen from Figure 12 the non-additivity of the $H_{\mathcal{D}}$ / interactions are positive and of the same order of magnitude as $|\mathcal{X}|$.

To conclude, we have seen that we can demonstrate the origin of some Hd/ effects in an extremely simple model. We do not consider this to be a realistic model. However, it is remarkable that the estimated values obtained from this model are of similar order of magnitude as in a real system.

4. The *Hol* interaction for real solutes in liquid water

We consider again a system of N water molecules in a volume V and temperature T. The solvation Helmholtz energy of a solute *s* is given by

$$\Delta A_s^* = -k_B T ln \ \langle exp \ [-\beta B_s] \rangle_0 \tag{4.1}$$

The solute s is assumed to have at least one arm along which it can form a HB with water molecules. We write the solute-solvent pair potential as

$$U(\boldsymbol{X}_{s},\boldsymbol{X}_{i}) = U^{**}(\boldsymbol{X}_{s},\boldsymbol{X}_{i}) + U^{HB}(am,\boldsymbol{X}_{i})$$
(4.2)

where $U^{HB}(arm, X_i)$ is the HB part of the interaction associated with the particular arm we are considering. U^{**} is defined in equation (4.2). The total binding energy can be written as

$$B_{s} = \sum_{i} U(\boldsymbol{X}_{s}, \boldsymbol{X}_{i}) = B_{s}^{**} + B_{s}^{HB}(am)$$

$$(4.3)$$

and the solvation Helmholtz energy of the solute may be split into two terms^{11,12}

$$\Delta A_{s}^{*} = \Delta A_{s}^{**} + \Delta A_{s}^{HB/**}$$
$$= -k_{B}Tm \left\{ \langle ep \left[-\beta B_{s}^{**} \right] \right\}_{0} \langle ep \left[-\beta B_{s}^{HB} \right] \rangle_{**} \right\}$$
(4.4)

The meaning of the two terms on the rhs of (4.4), correspond to splitting the solvation process of the solute *s* into two steps. First, we solvate the U^{**} part of the interaction, the corresponding Helmholtz energy of solvation is ΔA_{s}^{**} . Next, we solvate the HBing of the arm. The corresponding work is the conditional solvation Helmholtz energy of the arm given that all other parts of the interactions were already solvated.

A particular simple case is when the solute s is a hard sphere plus one arm. In this case U^{**} is simply the hard sphere pair potential. The corresponding solvation Helmholtz energy for this solute is

$$\Delta A_s^* = \Delta A_s^{HS} + \Delta A_s^{HB/HS}$$
$$= -k_B T \ln \langle exp \ [-\beta B_s^{HS}] \rangle_0 - k_B T \ln \langle exp \ [-\beta B_s^{HB}] \rangle_{HS}$$
(4.5)

In (4.5) the first term is the solvation Helmholtz energy of a hard sphere, or the cavity work. This is the analog of the work required to find an empty cell in equation (3.10). The second term is the *conditional* solvation Helmholtz energy of the HBing of an arm given that the hard part of the interaction has already been solvated.

As we have done in section 3, we focus on the change in the *conditional* solvation of one arm on each solute, or on the group of the protein.

Focusing on the second term of either eq. (4.4) or (4.5) we can simplify the average quantity as follows:

$$\langle exp \ [-\beta B_s^{HS}] \rangle_{HS} = \int exp \ [-\beta B_s^{HS}] P(\boldsymbol{X}^{V}|HS) d\boldsymbol{X}^{V}$$
 (4.6)

where $P(\mathbf{X}^{V}|\mathcal{B})$ is the conditional distribution of the configuration \mathbf{X}^{V} of the water molecules, given the HS at some specific location in the solvent. For the more general case, the conditional distribution is $P(\mathbf{X}^{V}|\mathcal{B})$ where the condition is the presence of the backbone (BB) of the protein.



The advantage of focusing on the conditional average in (4.6) is that now the integral can be split into two regions, in one the integration is over all configurations of water molecules such that there is one water molecule at the correct configuration near the arm to form a HB. The second includes all other configurations.

This can easily be seen by rewriting the average in (4.6) as

$$\langle exp \ [-\beta B_s^{HB}] \rangle_{HS} = \int_{-\infty}^{\infty} exp \ [-\beta \nu \] P(\nu) d\nu \tag{4.7}$$

where $P(\nu)$ is the distribution of the values of the binding energies of the solute, i.e.

$$P(\nu) = \int P(\boldsymbol{X}^{V} | HS) \delta[B_{s}(\boldsymbol{X}^{V}) - \nu] d\boldsymbol{X}^{V}$$
(4.8)

Thus, $P(\nu)d\nu$ is the probability of finding the binding energy of the solute between ν and $\nu + d\nu^{-11}$

A water molecule can form a HB with the arm only from a small region of the configurational space. Therefore, we rewrite the integral in (4.6) or (4.7) approximately in two terms

$$\langle exp \ [-\beta B_{s}^{HB}] \rangle_{HS} \approx exp \ [-\beta \varepsilon_{HB}] P_{HB} + (1 - P_{HB})$$
 (4.9)

Here we assume that the integrand can have only two values; when the arm is hydrogen bonded we have $\nu = \varepsilon_{IB}$, and when it is not hydrogen bonded $\nu = 0$. P_{IB} is the probability of finding a water molecule in the correct location and the correct orientation to form a HB with the arm. The probability of the complementary event is $(1 - P_{IB})$. As in section 3, the probability P_{IB} depends both on the density and on solvent-solvent interactions. This has important consequences regarding the entropy and the enthalpy changes associated with the H_{ID} interaction. This aspect will be discussed briefly in the next section.

$$\partial G_{2}^{Hdy} \approx -k_{B} T \ln \left\{ \frac{\exp\left[-\beta 2\varepsilon_{IB}\right] P_{IB}^{(2)} + \left(1 - P_{IB}^{(2)}\right)}{\left[\exp\left[-\beta \varepsilon_{IB}\right] P_{IB}^{(1)} + \left(1 - P_{IB}^{(2)}\right)\right]^{2}} \right\}$$
(4.10)

This is almost the same as equation (3.16) except for one important difference. In the model used in section 3 we have assumed that the probability of finding a water molecule in the correct configuration to form a HB with *one* arm, $P_{HB}^{(1)}$, is the same as the probability of finding a water molecule in the correct configuration to form *two* HBs with the two arms, $P_{HB}^{(2)}$. However, in reality these two probabilities are different. Using a very simple model for water (denoted BN3D in reference 11) we have estimated that the ratio of $P_{HB}^{(1)}/P_{HB}^{(2)}$ is about 360/20, i.e. $P_{HB}^{(2)}$ is smaller than $P_{HB}^{(1)}$ by a factor of about 18. This is due to the fact that in $P_{HB}^{(1)}$ we take into account the full rotation about the axis along which a single bond is formed, see Figure 13a. On the other hand only a small fraction of these configurations contribute to the formation of two HBs, Figure 13b.



Figure 13. Rotation about the axis connecting the centers of the water molecule and the solute do not affect the HB between the water and the solute's arm (a). On the other hand only a small fraction of these configurations are such that two HBs can be formed with two solutes (b).

Assuming again that $P_{BB}^{(1)}$ and $P_{BB}^{(2)}$ are small compared to 1 and that the factor $eqp [-\beta \varepsilon_{BB}]P_{BB}^{(1)}$ is large compared with one, we can get a rough estimate; for

 $\varepsilon_{HB} = -6kcal / mol$.

$$\delta G_2^{Hall} \approx -k_B T ln \quad \frac{P_{HB}^{(2)}}{P_{HB}^{(1)}} \approx -2 k c a l / m o l$$

(4.11)

and for $\varepsilon_{HB} = -6.5 kaal / mol$ $\delta G_2^{Hal} \approx -2.6 kaal / mol$



This is almost the same value we got almost 20 years ago¹ using a simple model for water and a more lengthy argument^{1,12}

5. Discussion and conclusions

Ever since a complete inventory of all possible solvent-induced effects in protein folding has been established it became clear that the role of the hydrophobic *HD* effect in protein folding was grossly exaggerated.^{2,6} The main reasons for this is that Kauzmann's model is based on the experimental values of the Gibbs energy of transferring of a non-polar solute from water into an organic liquid, Figure 1.

 $\Delta G_s^*(water \to organic \quad liquid \quad) = \Delta G_s^*(in \quad an \quad organic \quad liquid \quad) - \Delta G_s^*(in \quad water \quad) \tag{5.1}$

where on the rhs of (5.1) we have the solvation Gibbs energies of the solute s in an organic solvent and in water.

If the experimental values of ΔG_s^* (*transfer*) where applicable to proteins, then we would have obtained a large "driving force" for the protein folding process. Unfortunately, it was found that Kauzmann's model is inapplicable to HO groups attached to the protein backbone. Briefly, instead of ΔG_s^* (*water*) one must use the *conditional* solvation Gibbs energy of a HO group. Second, one does not need the solvation Gibbs energy of the HO group in an organic liquid. A more complete argument is given in references 11 and 12.

Yet, in spite of these findings the belief that the HpO effect dominates the "driving force" for protein folding still lingers. This is unfortunate because at the same time when it was shown that the HpO effect cannot contribute significantly to the driving force for protein folding, it was also shown that a host of HpO effects are much stronger than the corresponding HpO effects.

Recognizing the dominance of the *H*\$\$\$\$\$\$\$\$\$\$\$\$\$\$ effect has a profound impact on our ability to understanding two fundamental processes; the protein folding and protein-protein association.

There are essentially three problems that comprise the so-called Protein Folding Problem.

- 1. The relationship between a sequence and the native structure of the protein.
- 2. The factors that contribute to speed and guide the folding of proteins.
- 3. The factors that contribute to the stabilization of the native structure of proteins.

Problem 1 does not have a general solution, i.e. there exists no "code" that translates from a sequence of amino acids to a 3D structure. The second problem has a general answer. The most likely factors that speed and guide the folding process are the H_{a} forces.¹² Problem 3 also has a general answer. It was shown that both intramolecular HBs, as well as various H_{a} interactions are likely to be the main contributors to the stability of the native structure of the protein.¹²

The conjecture that there might be a "code," was probably inspired by Anfinsen's conclusion that the information for the native structure is contained in the sequence of the amino acids. However, Anfinsen was careful enough to add that this conclusion is valid for a specific environment, namely the temperature, pressure and solvent composition. Different environments might dictate different structures for the same sequence. Therefore, there can be no "code" that translates from a sequence to a structure. Equivalently, one can say that the information contained in the *system*, i.e. the sequence plus the environment, determines a unique structure, if such a structure exists. Clearly, in this formulation there will be infinite "codes" that translate from a specific "sequence plus the environment" into a 3D structure.

Although problem 1 has no general solution, the Hdt effect can contribute to the understanding of the reasons for this. Furthermore, inclusion of Hdt effects can help in the simulation of folding processes of specific sequences of amino acids.¹²

In this article I have endeavored to construct a "minimal" model to demonstrate the origin of the Hot effect. In this model we have eliminated all features of water which are not relevant to the process of Hot interactions. The most serious simplification of the model is the neglect of solvent-solvent interactions. As we have argued in sections 3 and 4, this assumption renders the solvability of the model, yet it does not affect the origin of the strong Hot effect.

It should be stressed however that this model is not appropriate for the study of the entropy and enthalpy changes associated with the processes studies in this article. The reason is that both the entropy and the enthalpy of the H_{H} interactions strongly depends on structural changes induced in the solvent, which is another way of expressing the effect of the H_{H} process on the average solvent-solvent interaction.^{11,12} These changes in the solvent-solvent interaction will not affect the value of the solvation Gibbs energy, nor that of the H Φ I interactions.^{12,17}

Another important aspect of the solute-induced interaction which we have demonstrated in this model is the non-additivity of the *Har* interaction. The sources of the non-additivity of the solvation Gibbs energy of any solute has been discussed in references 12 and 16.

Here, it is appropriate to point out that many models of protein folding use the assumption of pairwise additivity for the H_{D}^{O} interactions. However, since the pairwise additivity assumption cannot be used even as an approximation, it follows that any theory based on this assumption is invalidated. A further reason for the invalidation of the theory is the inclusion of H_{D}^{O} effect, but excluding the more important H_{D}^{O} effects. On the other hand, the H_{D}^{O} forces and interactions provide simple and intuitive answers to the problems of protein folding and protein-protein association.^{18,19,20}



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