



Kinetic studies of adsorption of Fe(III) from aqueous solution by untreated and alkali-treated rice straw

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ABSTRACT

Modifications of rice straws surfaces by sodium hydroxide (NaOH) were carried out in order to study the effects of this on the surface functional groups properties. Comparison was made between untreated and alkali-treated rice straws on the removal of Fe(III) from aqueous solution. In this study, four characterizations of raw rice straw have been conducted. The morphological characteristics by Scanning Electron Microscope (SEM), Energy Dispersive X-ray Analysis (EDXA) the functional group present in the rice straw by Fourier Transform Infrared (FTIR) spectroscopy and the X-ray Diffraction (XRD). The result of Scanning Electron Microscopy (SEM) also shows that rice straw is a porous material. Rice straw contains on -OH functional group that can bind with metal ions. To be able to enhance the sorption capacity of rice straw in metals, removing from waste water, alkali treatment should be done. This shows that rice straw can be used as adsorbent for ferric ions removing from wastewater. The effect of pH, sorption kinetics and isotherms were studied in batch experiments. The good correlation coefficient was obtained from pseudo second-order kinetic model, which agreed with conception as the rate-limiting mechanism. Sorption isotherm test showed that equilibrium sorption data were better represented by Temkin model. The highly efficient, low cost and the rapid uptake of Fe(III) by untreated (RS) in comparison with alkali-treated (MRS) rice straws, indicated that it could be an excellent alternative for the removal of ferric ions by sorption process.

Key words

Rice straw; Adsorption; Ferric; Kinetic; Thermodynamic

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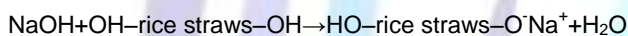
1. INTRODUCTION

Water pollution due to toxic heavy metals has been a major cause of concern. The industrial and domestic wastewater is responsible for causing several damages to the environment and adversely affecting the health of the people. Metals can be distinguished from other toxic pollutants, since they are non-biodegradable and can accumulate in living tissues, thus becoming concentrated throughout the food chain.

The main techniques, which have been utilized to reduce the heavy metal ion content of effluents, include lime precipitation, ion exchange, adsorption into activated carbon, membrane process, and electrolytic methods. All these methods are generally expensive. Therefore, numerous approaches have been studied for the development of low-cost adsorbents. Recently, [1,2] reviewed the technical feasibility of various low-cost adsorbents for heavy metal removal and concluded that the use of low-cost adsorbents may contribute to the sustainability of the surrounding environment and offer promising benefits for commercial purpose in the future.

Rice straw is an important agricultural crop residue generated as a by-product during dehusking at rice mills. For every ton of rice processed, rice straw production is estimated to be about 0.23 tons. A large amount of rice straw is burnt in situ, generating CO₂ and other forms of pollution. Thus the use of rice straw not only would provide a less costly sorbent to activated carbon or synthetic ion-exchanger as it is cheap and easily available, but will also save the environment from the above-mentioned pollution. Rice straw contains lots of silica. The organic compounds are mainly cellulose, hemicellulose and lignin. It was reported to be good sorbent for a variety of metals cation [3,4].

Alkali treatment of cellulosic fibers with sodium hydroxide (NaOH) is one of the chemical treatments methods that have been employed in order to improve the fiber-matrix interface bonding [5,6]. NaOH removes natural fats and waxes from the cellulose fiber surfaces thus, revealing chemically reactive functional groups like -OH. The removal of the surface impurities from the cellulose fibres also improves the surface roughness of the fibres or particles, thus opening more hydroxyl groups and other reactive functional groups on the surface [7]. NaOH may also react with accessible -OH groups according to the chemical reaction proposed as follow;



The reaction equations suggest reduction of -OH groups on the fiber surfaces, which is demonstrated as a decrease in -OH peak intensity in Fourier transform infrared (FTIR) spectra [8].

Rice straw was selected due to its local availability in abundance, chemical stability and insolubility in water. In the present study, the biosorption efficacy of rice straw for the abatement of Fe(III) ions from aqueous solutions has been investigated. The changes in the surface and adsorption properties of rice straw modified by NaOH were monitored by scanning electron microscopy (SEM) and attenuated total reflection (ATR) FTIR spectroscopy.

2. MATERIAL AND METHODS

2.1. Preparation of adsorbents:

2.1.a. Rice straw

The natural rice straw (RS) used in the present experiments was obtained from a market in El-Menoufia Governorate, Egypt. Its chemical compositions are shown in Table 1. The (RS) was thoroughly washed with a stream of distilled water to remove all dirt and then were dried at 110°C for 24 h to constant weight. The dried rice straws were stored in desiccators until used.

2.1.b. Modified rice straw

The modified rice straw (MRS) sample was prepared by alkali treatment. Alkali treatment was carried out by placing the RS sample in contact with NaOH (0.1 M), with constant stirring for 24 h. The liquid/solid ratio was 10 mL/g. The slurry was allowed to settle for 24 h. It was then filtered, washed OH⁻ free with distilled water, and dried at 110 °C for 24 h to constant weight. And it was ground and sieved. The particles 0.63 mm was selected

and preserved at room temperature in a sealed bottle.

2.2. Preparation of metal-solutions

The Fe(III) stock solution containing 1000 mg/L was prepared by dissolving ferric chloride (FeCl₃) (analytical reagent grade) in distilled water. Ferric working solutions in different concentrations was prepared by diluting the Fe(III) stock solution with distilled water.

2.3. Analytical technique:

The concentrations of the Fe(III) metal ions were performed using Flame Atomic Absorption Spectrophotometer (FAAS) Vario 6. Elements were determined using an air-acetylene flame.



3. RESULTS AND DISCUSSION

3.1. Characterization of adsorbents

3.1.1. Chemical composition

The chemical composition of the rice straw was determined at each stage of treatment and the data are summarized in Table 1.

Table 1 : Chemical Characterization of RS and MRS

No.	Chemical Characterization	Samples	
		RS	MRS
1	Moisture Content %	7	6.65
2	Ash Content %	6	13.3
3	Lignin Content %	12.5	13
4	Holocellulose %	75.5	72.8
5	Alpha Cellulose	56	61

3.1.2. FTIR spectra

The rice straw is constituted basically by cellulose, hemicellulose, lignin, extractives, water and mineral ash which is in large amount SiO₂. The lignin is promptly available to interact with cations, by firstly exchanging with protons and subsequently by chelating with the metallic ion.

The results of ATR-FTIR of the outer surfaces of NaOH treated and untreated rice straws are shown in Fig. 1(a,b). The term outer surface of rice straw means the outer part of the paddy grain before hulling and inner surface is that part of the paddy that houses the edible rice grain. A medium broad absorption band was found around the region of $\sim 3300\text{ cm}^{-1}$ for untreated rice straws. This band is due to stretching vibration of intermolecular hydrogen bonded $-\text{OH}$ groups in cellulose fibers [9].

After modification with NaOH the absorption band shifts to higher frequencies by $\sim 26\text{ cm}^{-1}$. This is an indication of the presence of free $-\text{OH}$ groups which do not take part in hydrogen bonding [9]. However, there was no evidence of a decrease in the intensities of these peaks possibly due to reaction of accessible $-\text{OH}$ groups and NaOH as proposed by earlier researchers [10].

A similar trend occurred to the other $-\text{OH}$ groups absorption peaks around $1626 \pm 4\text{ cm}^{-1}$, which shifted slightly to higher frequencies after treatment with NaOH suggesting an increase of free OH. Absorption represented by weak bands in the absorption region around $\sim 2925 \pm 1\text{ cm}^{-1}$ corresponds to the vibration of the carbon-hydrogen bonds superimposed onto $-\text{OH}$ broad band around $\sim 3300\text{ cm}^{-1}$.

Absorption vibration at $\sim 1738\text{ cm}^{-1}$ appearing on the outer surfaces of the untreated rice straws is due to the vibration of carbonyl from carboxylic groups in ester linkage as proposed by Trejo-O'Reil and Cavaille [11] or due to wax and natural fats. After treating rice straws with NaOH, this peak disappeared, which indicate that it might have been removed by this modification. Another carbonyl vibration occurred at $\sim 1539\text{ cm}^{-1}$ which could be a spurious band due to carboxylic group vibration or cell windows interaction. This peak disappeared also after increasing the concentration of NaOH.

A medium sharp peak around $\sim 1217\text{ cm}^{-1}$ due to vibration of silica bonds was observed on the outer surface of untreated rice straws. This peak disappeared on NaOH treated rice straws. Absence of this peak in NaOH treated rice straws is an indication of the possibility of sodium hydroxide to react with silica. A decrease in the silica vibration band at $786 \pm 5\text{ cm}^{-1}$ was observed as the concentration of NaOH increased. This is another evidence of the possibility of NaOH to react with silica on the outer surface of rice straws. It is therefore possible that part of the cellulose embedded with silica in the formation of silicon-cellulose membrane on the outer surface of rice straws [12] also degraded during disintegration of this membrane and silica. This may have contributed to lack of increase of the OH group bands on this surface after alkali treatment.

Fig. 1(a,b) shows the FTIR spectra of the inner surfaces of untreated and NaOH treated rice straws. A broad peak in the region of $\sim 3273.5 \pm 57\text{ cm}^{-1}$ in untreated rice straws is due to hydrogen bonded $-\text{OH}$ in cellulose fibers. There is also a slight shift of the $-\text{OH}$ peak to high frequencies in NaOH treated rice straws. The shift of this peak by about 49 cm^{-1} to high frequencies suggests presence of free OH. The intensity of this peak is relatively weak for untreated rice straws, but increases progressively with the increase in the concentration of NaOH. The reason behind this could probably be due to the removal of surface impurities from the surface of rice straws thus exposing more reactive $-\text{OH}$ groups on these surfaces, which were detected by ATR-FTIR.

Weak absorption bands around $\sim 2919.5 \pm 0.5\text{ cm}^{-1}$ due to C-H vibration appears immediately after treatment of rice straws with NaOH. This is another evidence of the removal of impurities from the inner surface of rice straws by NaOH. The absorption band around $\sim 1614 \pm 19\text{ cm}^{-1}$ by a medium, stretching bond may be due to $-\text{OH}$ vibration whose intensity increased after treatment. The absorption peak around $1201\text{--}1156\text{ cm}^{-1}$ occurs in the vibration range of silica bonds.

This band is common to all the spectra with no evidence of significant change in intensity after treatment. Another absorption corresponding to the absorption frequency of silica bonds appearing around 790 cm^{-1} for untreated rice straws disappeared in NaOH treated rice straws except for the one treated with 0.1M NaOH. However, no evidence of silica composition on the inner surface of rice straws has been reported in previous studies [12].

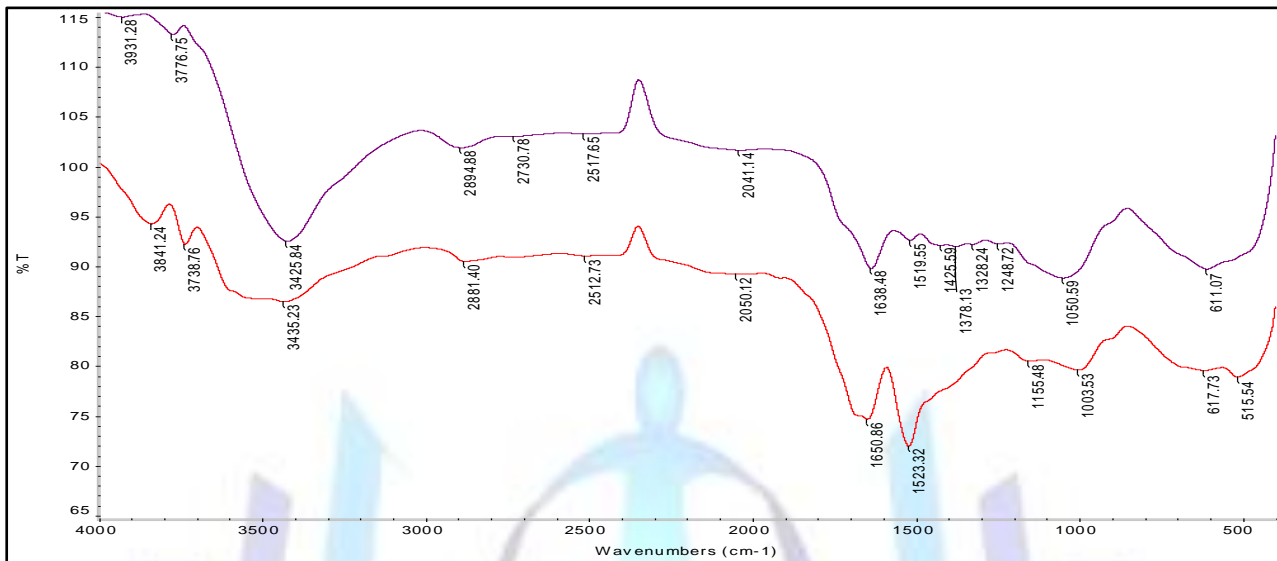


Figure 1 FTIR spectra for (a) RS and (b) MRS before adsorption.

3.1.3. Scanning electron

Scanning electron (SEM) micrographs of untreated and treated rice straws are shown in Fig. 2(a,b). As shown in Fig. 2(a,b) the surface roughness of the outer and inner surfaces of rice straws change significantly after alkali treated with NaOH. Changes started to be substantial when the rice straws were treated with NaOH. These include wearing of asperities on the outer surfaces and particle cracking which suggests the weakening of the rice straws due to increase in brittleness. The result of Scanning Electron Microscopy (SEM) also shows that rice straw is a porous material [13].

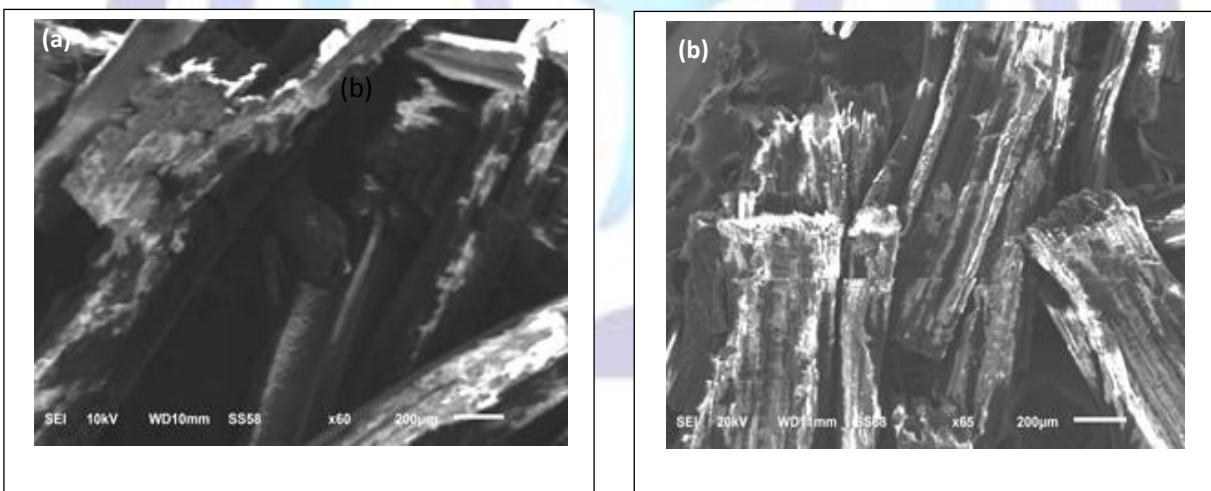


Figure 2 SEM image for (a) RS and (b) MRS before adsorption.

3.1.4. EDXA Spectra

Fig. 3(a,b) shows the EDXA spectra of RS and MRS adsorbents before and after loading with Fe(III) respectively. Fig. 3(a,b), indicates the presence of major constituents – carbon and oxygen in the two samples adsorbents. Comparing the spectra of the MRS loaded with Fe(III) with that of unloaded one, the cobalt peak could be observed. It was suggested that heavy metals including Fe(III) had been adsorbed on the surface of MRS successfully. Moreover, after loading with heavy metal, a distinct increase of silica peak intensity could be found. This phenomenon might be derived from the alkali treatment [14].

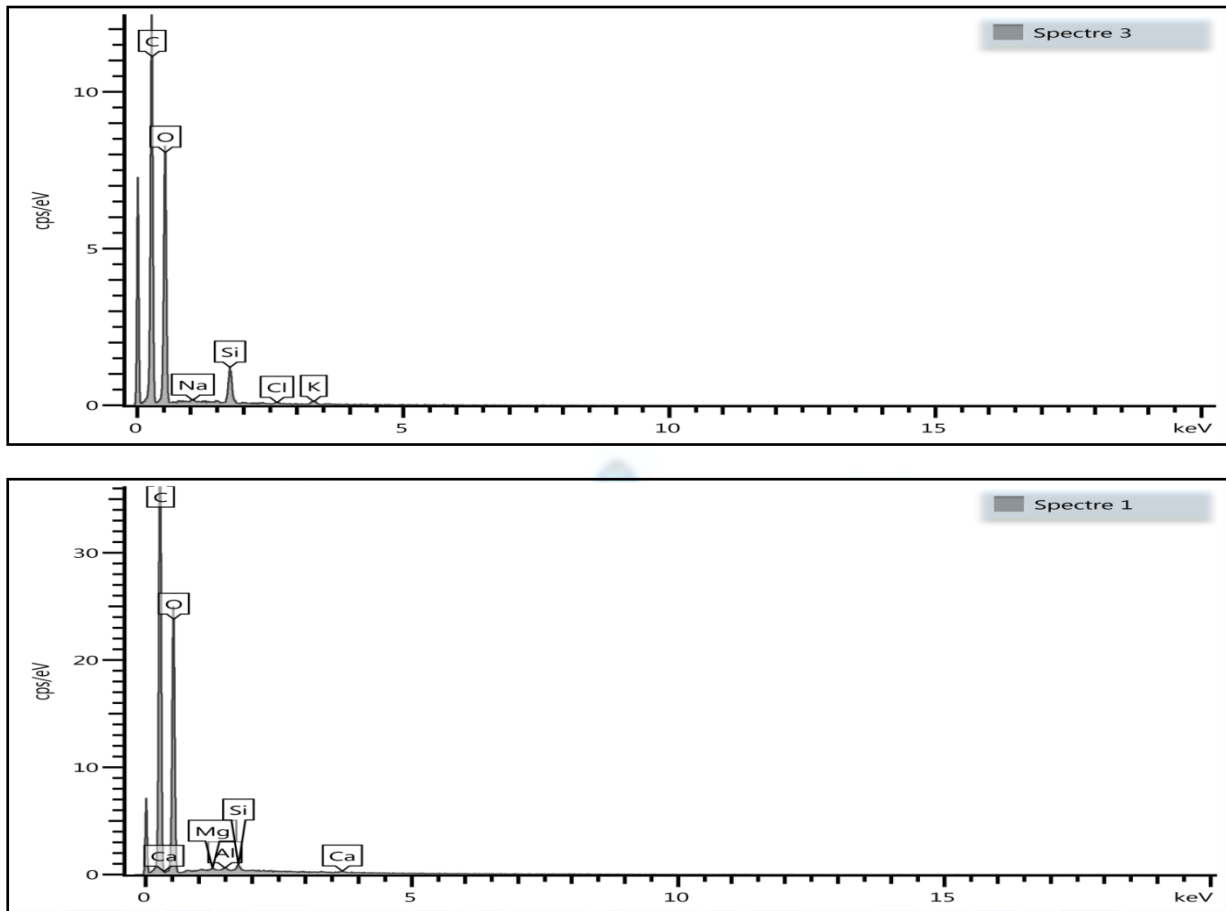


Figure 3 EDXA for (a) RS and (b) MRS before adsorption.

3.1.5. X-ray diffraction

Fig. 4(a,b) shows the X-ray diffraction patterns of RS and MRS biosorbents powder. Diffraction peaks corresponding to crystallinity were not observed, whereas, amorphous form is identified in the biosorbent. The amorphous nature of the biosorbents suggested that the metal ion could more easily penetrate into the surface of the two biosorbents on the amorphous. Optimization of Fe(III) biosorption by chemically modified rice straw [15].

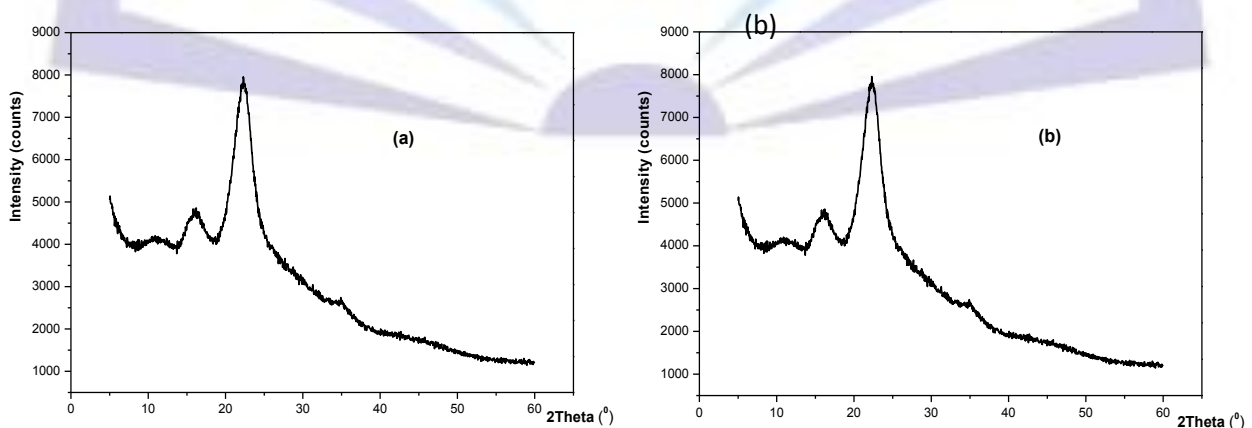


Figure 4 XRD for (a) RS and (b) MRS before adsorption.

3.2. Effect of adsorbent dosage on adsorption

Fig. 5(a,b) shows the removal of ferric as a function of adsorbent dosage using RS and MRS in aqueous solution. The adsorbent dosage varied from 0.1 to 1.0 g and equilibrated for ≈ 5 h. It is clear from fig.5(a,b) that for the maximum removal percentage of 85.9 and 53.3% of ferric requires a maximum RS and MRS dosage of 1.0 g for, respectively. The data clearly shows that all the adsorbents have a high level of performance in terms of the removal of ferric. The observed differences may be due to the high adsorption capacity of RS. It may be concluded that by increasing the adsorbent dose the removal efficiency increases but adsorption density decreases. The decrease in adsorption density can be attributed to the fact that some of the adsorption sites remain unsaturated during the adsorption process; whereas the number of available adsorption sites increases by an increase in adsorbent and these results in an increase in removal efficiency.

As expected, the equilibrium concentration decreases with increasing adsorbent doses for a given initial ferric concentration, because for a fixed initial solute concentration, increasing the adsorbent doses provides a greater surface area or adsorption sites [16].

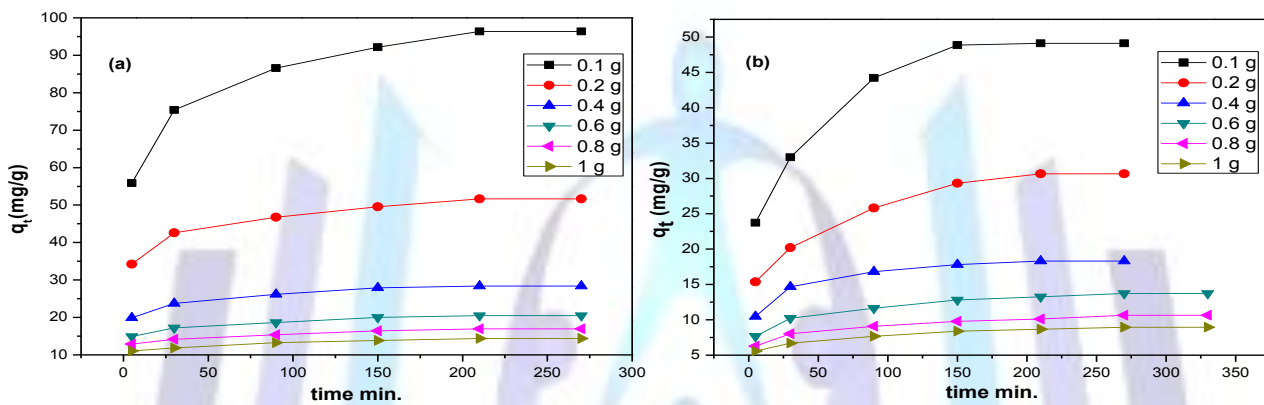


Figure 5 The effect of amount of (a) RS and (b) MRS on the removal of Fe(III) at 28 °C.

3.3. Effect of initial ferric concentration

The Fe(III) adsorption capacity increased with the Fe(III) equilibrium concentration increasing from 111.7 to 781.9 mg/L. This capacity of the RS was ranged from 23.1 to 40 mg/g and from 13 to 22.6 mg/g, of MRS, respectively. On the other hand, we can observe that, with an increase of the Fe(III) equilibrium concentration, the removal percentage of ferric, show an opposite trend where, the removal percentage was decreased from 78.8 – to 40.9% and from 93.1 to 23.1%. The results of the Fe(III) adsorption experiments are shown in Fig. 6. Actually, as the initial concentrations of Fe(III) increased, the driving force became higher as well, the accessibility of the heavy metal ions to the binding sites of the MRS is relatively high with increased initial concentration, the ions exchange frequently and the uptake of heavy metals becomes more and more [17].

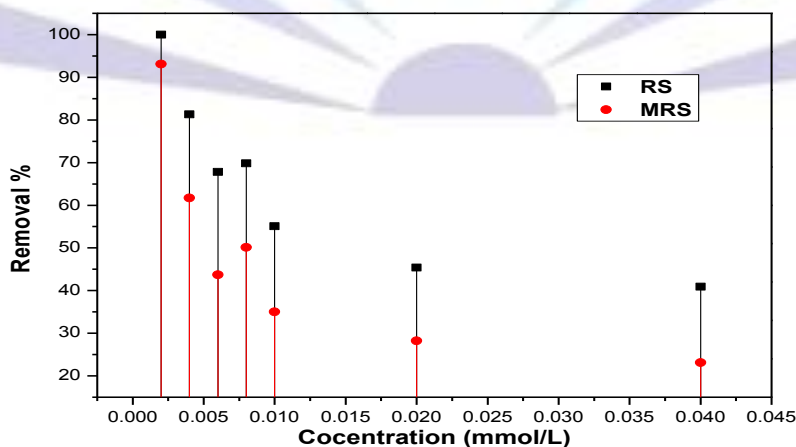


Figure 6 The effect of different concentration of Fe(III) on the adsorption.

3.4. Effect of pH on Fe(III) adsorption

The effect of pH on the Fe(III) adsorption on the two adsorbents for a pH between 1.0 and 2.9 is presented in fig.7. It can be found that the removal efficiency, increased with increase pH for both adsorbents. The uptake of Fe(III) by RS and MRS increased as the pH increased from 1.0 to 2.5. At higher pH value, 2.9 the removal efficiency decreased for both adsorbents. Although a maximum uptake was noted at a pH of 2.5, as the pH of the solution increased to >2.5 Fe(III) started to precipitate out from the solution. Therefore, the increased capacity of adsorption at pH = 2.5 may be a combination of both adsorption and precipitation on the surface of the adsorbents. It is considered that adsorbents had a maximum adsorption capacity at a pH=2.5, if the precipitated amount is not considered in the calculation. Therefore, the optimum pH for Fe(III) adsorption is 2.5.

The pH of the aqueous solution is an important variable that influences the adsorption of anions and cations at the solid–liquid interfaces. As can be seen from Fig. 7, the pH value of the ferric solution plays an important role in the whole adsorption process and particularly on the adsorption capacity. The Fe(III) adsorption on the two adsorbents tends to increase with the increase of pH. This is likely attributed to the fact that a lower pH value causes the surface to carry more positively charges and thus would more significantly repulse the positively charged species in solution. Therefore, the lower adsorption of Fe(III) at lower pH values resulted from an increased repulsion between the more positively charged Fe(III) species and positively charged surface sites. Furthermore, at lower pH, H⁺ ions compete with Fe(III) ions to the surface binding-sites of the adsorbent [18-20].

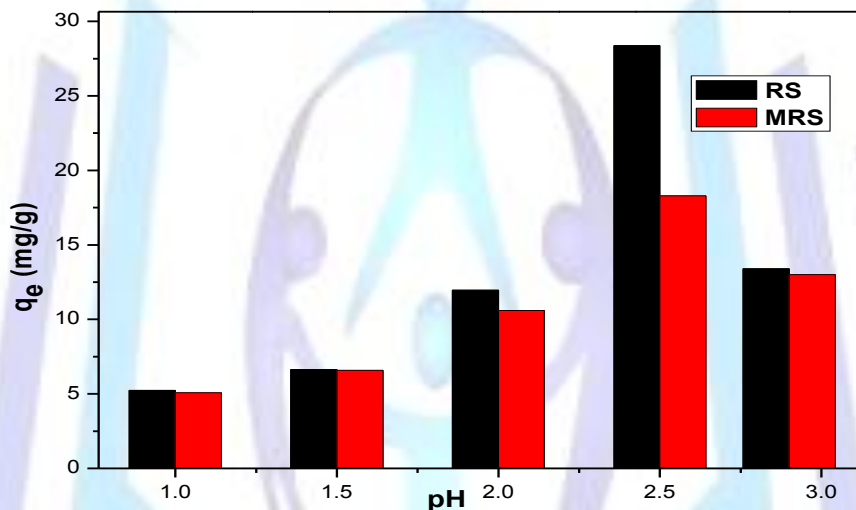


Figure 7 The effect of pH on the removal of Fe(III) at 28 °C.

3.5. The distribution ratio (D)

Distribution ratio D for ferric ions was determined by the batch method at different temperature systems (301, 313 and 323K). The distribution ratio, D , is defined as the ratio of metal ion concentration on the adsorbent to that in the aqueous solution and can be used as a valuable tool to study Fe(III) ion mobility. The distribution ratio D is defined by the following relationship [21,22]:

$$K_d = \frac{(I-F)}{500 \text{ mg}} \times \frac{50 \text{ ml}}{F}$$

Where I is the volume of EDTA used before treatment of metal ion-exchanger. F is the volume of EDTA consumed by metal ion left in solution phase.

Various portions of (0.4g each) the adsorbent were taken in Erlenmeyer flasks and mixed with 50 ml of metal ion solution in the aqueous medium and subsequently shaken for 24 h in temperatures controlled shaker at 301, 313 and 323K to attain the equilibrium. Fig. 8 shows that the distribution ratio(D) values increase with the increase in temperatures of ferric solutions from 301 to 313K then decrease at higher temperature (323K). However, the distribution ratio of Fe(III) between ferric solution and MRS show a gradual increases at increase of temperature. High values of the distribution ratio (as in aqueous-RS systems at low temperature), indicate that the metal has been retained by the solid phase through sorption reactions, while lower values of D (as in aqueous-MRS systems at low temperature), indicate that a large fraction of the metal remains in solution. The rapid metal sorption has significant practical importance, as this will facilitate with the small amount of resin to ensure efficiency and economy.

According to the above results of the Fe(III) adsorption experiments, the RS had higher adsorption capacities than the MRS. It was believed that the surface structural changes of the material play the most important role in the adsorption capacities of the Fe(III). When the RS sample was treated with 0.1M NaOH, the surface structure of the RS was changed, which can be seen from the FTIR spectra of the MRS.

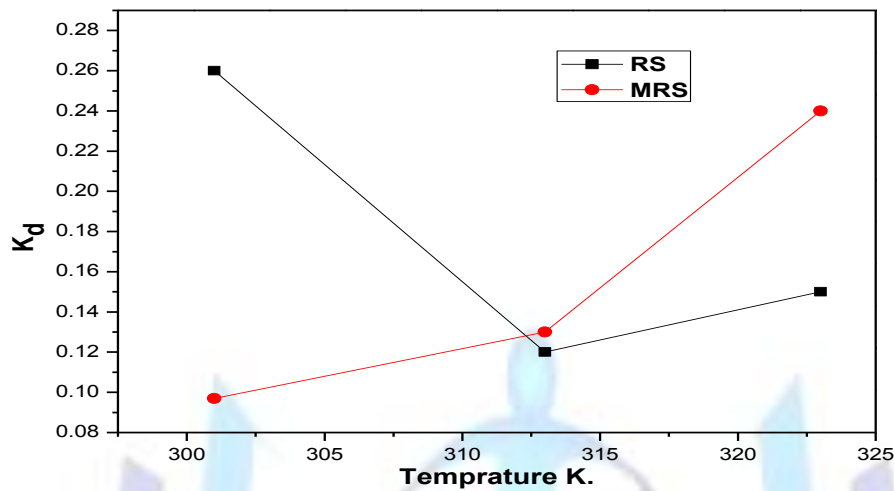


figure 8 The effect of distribution coefficient on adsorption of Fe(III).

3.6. Adsorption isotherms

The equilibrium adsorption isotherms of ferric ions were determined at three different temperatures (301, 313 and 323 K). The experimental data were fitted to linear equations of Freundlich [23,24], Langmuir [25,26], Temkin [27,28] and Dubinin and Radushkevich [29,30], isotherm models.

Two typical isotherms, as described below in Eq. (1) and (2), were used for fitting the experimental data:

$$\log q_e = \log K_F + \frac{1}{n} \log C_e \text{ ----- 1}$$

$$\frac{C_e}{q_e} = \frac{1}{K_L Q_{max}} + \frac{C_e}{Q_{max}} \text{ ----- 2}$$

Where q_e is the amount adsorbed at equilibrium (mg/g), and C_e is the equilibrium concentration (mg/L). The other parameters are different isotherm constants, which can be determined by the correlation coefficient of the experimental data. The value of (n) is a characteristic constant measure of intensity of sorption. The values of (n) computed from the slope of the plots of Freundlich equation (Fig. not shown) (1.5 and 1.3 for ferric on both RS and MRS, respectively) and the results of k_F (mg/g) estimated from the intercept of the linear plots. k_F was also 601.3 and 1174.9 of both RS and MRS, respectively, indicate better sorption at the experimental conditions.

A dimensionless constant, separation factor, R_L describes the type ($R_L = 0$ irreversible, R_L between 0 and 1 favourable and unfavourable ($R_L > 1$) of Langmuir isotherm, which is an essential characteristic of Langmuir isotherm and may be calculated in the temperature range (301-323K) for cobalt ion, by employing the relationship;

$$R_L = 1 / (1 + K_L C_i)$$

Where ' K_L ' is the Langmuir constant ($L \text{ mol}^{-1}$) and C_i is the initial concentration of sorbate (mg/g). In linear Plots of C_e/q_e versus C_e (Fig. not shown), the values of Q_{max} are analyzed from the slope of the linear plots and its values are 188 and 12.2 mg/g of RS and MRS, respectively, whereas the values of ' K_L ' (1.4×10^{-3} and 0.016 mol L^{-1} of RS and MRS, respectively) for Fe(III) are computed from the intercepts of the plots. The values of R_L are 0.68 and 0.16 of RS and MRS, respectively; assign a highly favorable sorption at whole solution temperatures studied herein.

Dubinin–Radushkevich (D–R) isotherm is another adsorption model isotherm applied in the linearized form of the equation(3) as,

$$\ln q_e = \ln X_m - \beta \epsilon^2 \text{ ----- 3}$$

Where q_e is in (mg/g) (described earlier), X_m ((mg/g)) represents the maximum sorption capacity of sorbent and β ($\text{kJ}^2 \text{ mol}^{-2}$) is a constant with dimensions of energy. The Polanyi sorption potential ϵ , which is the amount of energy required to pull a sorbed molecule from its sorption site to infinity may be evaluated by using relationship equation (4) as:

$$\epsilon = RT \ln(1 + 1/C_e) \text{ ----- 4}$$

Where ' R ' is a gas constant in $\text{kJ mol}^{-1} \text{ K}^{-1}$, ' T ' is the temperature in Kelvin, C_e (mg/g) is as mentioned earlier. The plots of $\ln q_e$ versus ϵ^2 yield a poor coefficients (≈ 97 and 92) and the results of X_m computed from the slope and intercept of the



respective plots (Fig. not shown) (21.1 and 15.8 (mg/g). The values of sorption energy, E_{was} 3.4 and 21.9 kJ mol⁻¹ of both RS and MRS, respectively, for Fe(III) are calculated[31] using the relationship in equation (5)as:

$$E_{D-R} = \frac{1}{\sqrt{-2\beta}} \text{-----} 5$$

These values indicate physical and chemical processes for ferric-RS and -MRS systems, respectively. The Temkin isotherm has been used in the following equation (6):

$$q_e = B_T \ln A_T + B_T \ln C_e \text{-----} 6$$

Where R is gas constant (8.314 J mol⁻¹ K⁻¹), T is temperature (K), A_T is the equilibrium binding constant (L g⁻¹) corresponding to the maximum binding energy, and constant $B_T = (RT/b_T)$ is related to the heat of adsorption. A plot of q_e versus $\ln C_e$ (Fig. not shown) is used to calculate the Temkin isotherm constants A_T and B_T .

The values of B_T (J/mol) are analysed from the slope of the linear plots and its values are 17.0 and 18.4 mg/g of RS and MRS, respectively, whereas the values of A_T (1.7×10⁻³ and 1.97×10⁻³L/g) of RS and MRS, respectively, for Fe(III) are computed from the intercepts of the plots.

From our results we can concluded that the experimental data of Fe(III) adsorption on RS sample could be well fitted by the isotherms. Clearly, the temkin equation provided better fitting in terms of R^2 values (0.99082). However, the fitting order of the different adsorption models according to R^2 was as follows; Temkin (0.99082) > Langmuir (0.99037) > Freundlich (0.98752) > D – R (0.97017). on the other hand, the adsorption of Fe(III) on MRS provided better fitting of the temkin equation in terms of R^2 values (0.99556). However, the fitting order of the different adsorption models according to R^2 was as follows; Temkin (0.99556) > Langmuir (0.97732) > Freundlich (0.96688) > D – R (0.92946).

3.7. Adsorption kinetics

Kinetic studies were carried out using different models, namely, pseudo-first order [32,33], pseudo-second order [34,35], Elovich [36] and Fickian diffusion intraparticle [37,38] models to analyze the experimental data.

The results of the Fe(III) adsorption kinetic experiments at the 28°C show that the majority of Fe(III) adsorption on the adsorbents was completed in 3-4 h. For example, after 210 min of adsorption, the Fe(III) adsorbed on the RS and MRS was, respectively, 67.8% and 43.7% of that at equilibration time. And the removal percentage of ferric from the solutions by RS was higher than the MRS sample, which can be attributed to the surface structural changes of the material.

The adsorption kinetics, demonstrating the solute uptake rate, is one of the most important characteristics, which represents the adsorption efficiency of the samples. The Fe(III) adsorption kinetic data ((Fig. not shown)) were fitted with pseudo-first-order rate equation of Lagergren and pseudo-second-order rate equation of linear equations.

The pseudo-first-order kinetic model is given equation (7) as:

$$\log(q_e - q_t) = \log q_{e,1} - k_1 t \text{-----} 7$$

The pseudo-second-order equation is expressed equation (8,9)as:

$$\frac{t}{q_t} = \frac{1}{k_2 q_{e,2}^2} + \frac{t}{q_{e,2}} \text{-----} 8$$

$$h = k_2 q_{e,2}^2 \text{-----} 9$$

Where k_1 is the Lagergren adsorption rate constant (min⁻¹) and k_2 is the pseudo-second-order adsorption rate constant (g/(mg h)); q_e and q_t are the amounts of Fe(III) absorbed (mg/g) at equilibrium and time t , respectively. Based on R^2 obtained, the kinetics of Fe(III) adsorption on the RS (0.99862) and MRS(0.99873) can be satisfactorily described by the pseudo-second-order equations at 301K as in table 2.

Therefore, the fitting curves resulting from both equations are plotted (Fig. not shown). The high applicability of the pseudo-second-order equation for the present kinetic data is generally in agreement with other researchers' results that the pseudo-second-order equation was able to describe properly the kinetics of Fe(III) adsorption [39,40]. On the other hand, when the temperature was increased, the initial adsorption rate h (mmol/(g min)) of RS and MRS decreased from 5.4 to 0.78 and from 2.6 to 1.4 mg/(g min). The value (h) for RS was higher than that of MRS at low temperature, suggesting that RS possesses the fastest kinetics among two adsorbents as in table 2.

For Fickian diffusion law, all the correlation coefficients were relatively low and the intercept of plots revealed obvious boundary layer effect (Fig. not shown). Larger intercept means a greater contribution of surface adsorption as the rate-controlling step. In addition, it was essential for the plots of q_t versus $t^{0.5}$ to go through the origin if the intra-particle diffusion was the sole rate-limiting step. However, all the linear portions did not pass through the origin (all intercepts were in the range of 0.93914 - 0.97118 and 0.87999 - 0.95641 of RS and MRS, respectively), indicating that intra-particle diffusion maybe not only the rate-controlling factor [41]. This was further evidence indicating that the active sites of the two adsorbents are mainly distributed on the external surface. The adsorption rate of RS was faster than MRS because of its higher external surface area. Therefore, the external surface of the adsorbent was the key factor in the rate-controlling as in Table 2.



Table 2 : Kinetic parameters for (6 mmol/L) of Fe(III) on on RS and MRS in aqueous solution.

Rice Straw	Temp.K	Pseudo first-order model			Pseudo second-order model				Intraparticle diffusion model		
		q _{e,1,cal} (mg/g)	K ₁ (min ⁻¹)	R ²	q _{e,2,cal} (mg/g)	K ₂ (g/mg min)	h (mg/g min)	R ²	K _{int} mg/g min ^{-0.5}	C (mg/g)	R ²
RS	301	9.50	0.0194	0.96467	28.89	6.4×10 ⁻³	5.4	0.99862	0.686	19.197	0.93914
	313	16.22	0.0162	0.90001	25.79	9.2×10 ⁻³	0.16	0.95001	0.934	5.987	0.96538
	323	17.22	0.0171	0.916825	28.99	9.3×10 ⁻³	0.78	0.95498	0.954	8.492	0.97118
MRS	301	7.41	0.0181	0.98212	18.74	7.4×10 ⁻³	2.6	0.99873	0.612	10.222	0.87999
	313	12.02	0.0170	0.98829	22.07	3.7×10 ⁻³	1.8	0.99778	0.756	10.107	0.92768
	323	21.38	0.0173	0.95086	29.27	1.6×10 ⁻³	1.4	0.99299	1.240	8.789	0.95641

3.8. Effect of temperature

The equilibrium removal of Fe(III) ions as a function of temperature, for experiments conducted at constant concentrations of Fe(III) equal to 335.01 mg/L. The adsorption of Fe(III) onto the surface of both RS and MRS place quickly regarding the temperature (301–323 K). On the other hand, enhancement of the adsorption capacity of the MRS at higher temperatures may be attributed to the activation of the adsorbing surface and increase in the mobility of metal ions. Also, this fact demonstrated an endothermic biosorption process.

The removal percentage of cobalt onto the RS and MRS adsorbents (Fig. not shown) was increased with increase of temperatures at low 301 and 313K then decrease at higher temperature (323K) for RS. However, the removal percentage by MRS was increases with increase in temperature from 301 to 323 K. This may be due to the formation of new active sites in the adsorbents to increase in temperature, activation of the adsorbing surface and increase in the mobility of metal ions. An increase of adsorption capacities of Fe(III) on the two adsorbents as the temperature increased, indicating also an endothermic process and a possible type of chemical adsorption mechanism occurs.

3.9. Thermodynamics of sorption

Thermodynamic parameters, enthalpy ΔH (kJ mol⁻¹), entropy ΔS (J mol⁻¹ K⁻¹) and standard free energy of activation ΔG (kJ mol⁻¹) were investigated in the range of 301–323 K under the optimized conditions chosen by applying the equations(10,11) as:

$$\ln K_d = \frac{\Delta S}{R} - \frac{\Delta H}{RT} \text{-----10}$$

$$\Delta G = \Delta H - T\Delta S \text{----- 11}$$

Where 'R' is a gas constant, 'T' is the temperature in Kelvin. The plots of $\ln K_d$ versus $1/T$ (K⁻¹) are linear throughout the investigation and the values of ΔH and ΔS are computed from the respective slopes and intercepts of the plots. The calculated thermodynamic parameters are presented in Table 3. ΔH° values were negative, demonstrating an exothermic process in ferric-RS system while the ΔH° values were positive, demonstrating an endothermic process in ferric-MRS system. The positive ΔG° values accompanied by the negative ΔS° suggested that the sorption reactions are nonspontaneous with a low affinity and presence of high energy barrier in case of aqueous ferric-RS system.

On the other hand, the negative ΔG° values and the positive ΔS° suggested that the sorption reactions are spontaneous with a high affinity and alower energy barrier for ferric adsorption using MRS. This is may be due to changes in the surface functional groups of the adsorbent [42,43].



The positive enthalpy change (ΔH°) values for the metal ions adsorption reaction as in table 3 indicate the endothermic nature of the present reaction. ΔH° values obtained from adsorption of Fe(III) onto the MRS are lower than that onto RS. This result gives clear evidence that the interactions between Fe(III) and the surface groups of the RS may be weaker than that of the surface groups of the MRS. The low value of ΔH° ($< 40 \text{ kJ mol}^{-1}$) for Fe(III) onto both adsorbents indicated that the adsorption process occurs mainly through a physical means. On the other hand, the negative values of E_a (-13.88 kJ/mol) and ΔH° indicate the presence of high an energy barrier in the adsorption process in case of RS, while, the positive values of E_a (20.3 kJ/mol) and ΔH° indicate the presence of low an energy barrier in the adsorption process in case of MRS[44].

The positive values for these parameters are quite common because the activated complex in the transition state is in an excited form. The positive entropy change (ΔS°) for this reaction (Table 3) has also indicated the increase in the number of species at the solid-liquid interface and, hence the randomness in the interface which is presumably due to the release of aqua molecules when aquoted metal ion is adsorbed on the surface of the adsorbent and significant changes occur in the internal structure of the adsorbent through the adsorption of the metal ions onto the resin[45].

In order to further support the assertion that the adsorption is the predominant mechanism, the values of the activation energy (E_a) and sticking probability (S^*) were estimated from the experimental data. They were calculated using a modified Arrhenius type equation related to surface coverage as expressed in equations (12, 13) as:

$$\theta = 1 - \frac{C_e}{C_0} \text{-----} 12$$

$$S^* = (1 - \theta) \exp\left(-\frac{E_a}{RT}\right) \text{-----} 13$$

The sticking probability, S^* , is a function of the adsorbate/adsorbent system under consideration and is dependent on the temperature of the system. The parameter S^* indicates the measure of the potential of an adsorbate to remain on the adsorbent indefinitely. It can be expressed as in Table 3.

The effect of temperature on the sticking probability was evaluated throughout the temperature range from 301 to 323 K by calculating the surface coverage at the various temperatures. Table 3 also indicated that the values of $S^* \leq 1$ for MRS, hence the sticking probability of the Fe(III) ion onto RS are very high. However, the values of $S^* > 1$ for RS, hence the sticking probability of the Fe(III) ion onto RS are very low [46].

Table 3 : Thermodynamic parameters for (6 mmol/L) of Fe(III) on RS and MRS in aqueous solution.

Rice Straw	Temp.K	ΔG (kJ/mol)	ΔS (J/mol k)	ΔH (KJ/mol)	S^*	E_a (KJ/mol)
RS	301	26.6	-88.40	-23.00	88.17	-13.88
	313	27.6	-	-	-	-
	323	28.5	-	-	-	-
MRS	301	-30.1	100.27	36.38	1.86×10^{-4}	20.30
	313	-31.3	-	-	-	-
	323	-32.3	-	-	-	-

REFERENCES

- [1] Ihsanullah, Aamir Abbas, Adnan M. Al-Amer, Tahar Laoui, Mohammed J. Al-Marri, Mustafa S. Nasser, Majeda Khraishah, Muataz Ali Atieh; Separation and Purification Technology, 2016, 157, 141–161.
- [2] Yunhai Wu, Yiang Fan, Meili Zhang, Zhu Ming, Shengxin Yang, Aynigar Arkin, Peng Fang; Biochemical Engineering Journal, Part A, 15 January 2016, 105, 27–35
- [3] Jong-Hwan Park, Yong Sik Ok, Seong-Heon Kim, Ju-Sik Cho, Jong-Soo Heo Ronald D. Delaune, Dong-Cheol Seo; Chemosphere Volume , January 2016, 142, 77–83
- [4] Upendra Kumar, Manas Bandyopadhyay; Bioresource Technology, 2006, 97, 104–109.
- [5] Alireza Bazargan, Tesfalet Gebreegziabher, Chi-Wai Hui, Gordon McKay; biomass and bioenergy in press, 2014, 1-8.
- [6] A.K. Mohanty, M.A. Khan, G. Hinrichsen; Compos Sci Technol, 2000, 60, pp. 1115–1124.
- [7] J. George, M.S. Sreekala, S. Thomas; Polymer Eng Sci, 2001, 41 (9), pp. 1471–1556.
- [8] M.S. Sreekala, S. Thomas; Compos Sci Technol, 2003, 63, pp. 861–869.
- [9] S. Wong, R. Shanks, A. Hodzic; Compos Sci Technol, 2004, 64, pp. 1321–1330.
- [10] M.S. Sreekala, S. Thomas; Compos Sci Technol, 2003, 63, pp. 861–869.
- [11] J.-T. Trejo-O'Reil, J.-Y. Cavaille; Cellulose, 1997, 4, pp. 305–320.



- [12] S. Yoshida, Y. Ohnishi, K. Kitaishi; The chemical forms, mobility and deposition of silicon in rice plant; *Soil Sci Plant Nutr* (Tokyo), 1962, 8 (15).
- [13] Hengpeng Ye, Qing Zhu, Dongyun Du; *Bioresource Technology*, 2010,101, 5175–5179.
- [14] Yuan Feng, Ji-Lai Gong, Guang-Ming Zeng, Qiu-Ya Niu, Hui-Ying Zhang, Cheng-Gang Niu, Jiu-Hua Deng, Ming Yan; *Chem. Eng. J.*, 2010, 162, 487–494.
- [15] D. Harikishore Kumar Reddy, K. Sessaiah, A.V.R. Reddy, S.M. Lee; *Carbohydrate Polymers*, 2012, 88, 1077– 1086.
- [16] Ho, Y.S., Wase, D.A.J., Forster, C.F.; *Water Res.*,1995, 29,1327–1332.
- [17] Yunhai Wu, Yiang Fan, Meili Zhang, Zhu Ming, Shengxin Yang, Aynigar Arkin, Peng Fang; *Biochemical Engineering Journal*, 2016, 105, 27–35.
- [18] L. Largette, T. Brudey, T. Tant, P. Couespel Dumesnil, P. Lodewyckx; *Microporous and Mesoporous Materials*, 2016, 219, 265-275.
- [19] Congcong S., C. Chen, T. Wen, Z. Zhao, X. Wang, A. Xu; *J.Colloid and Interf. Sci.*, 2015, 456, pp. 7-14.
- [20] Lichun F., C. Shuang, F. Liu, A. Li, Y. Li, Y. Zhou, H. Song; *J.Hazard. Mater.*, 2014, 272, pp. 102-111.
- [21] S.A. Nabi, Mohammad Shahadat, Rani Bushra, A.H. Shalla, A. Azam; *Colloids and Surfaces B: Biointerfaces*, 2011, 87, 122–128.
- [22] Asif A. K., S. Shaheen; *Solid State Sciences*, 2013, 16,158-167.
- [23] Freundlich, H., *J. Am.; Chem. Soc.*,1918, 40, 1361-1403.
- [24] Yanping Zhu, Naiyun Gao*, Qiongfang Wang, Xingya Wei; *Colloids and Surfaces A: Physicochem. Eng. Aspects*, 2015, 468, 114–121.
- [25] Langmuir I.; *J. Am. Chem. Soc.*,1918, 40, 1361–403.
- [26] Mary Jenish Barnabas, Surendran Parambadath, Aneesh Mathew, Sung Soo Park, AjayanVinu, Chang-Sik Ha; *Journal of Solid State Chemistry*, 2016, 233, 133–142.
- [27] Temkin I.M., V. Pyzhev; *Acta Physicochem. SSR*, 1940, 12, pp. 217–222.
- [28] Guiyin Zhou, Jinming Luo, Chengbin Liu, Lin Chu, Jianhong Ma, Yanhong Tang, Zebing Zeng, Shenglian Luo; *Water Research*, 2016, 89, 151-160.
- [29] Dubinin, M.M., L.V. Radushkevich; *Chem. Zentr*, 1947, 1, pp. 875–889.
- [30] Hong-Tao Fan, Wen Sun, Bing Jiang, Qing-Jie Wang, Da-Wu Li, Cong-Cong Huang, Kang-Jun Wang, Zhi-Gang Zhang, Wen-Xiu Li; *Chemical Engineering Journal*, 2016, 286, 128–138.
- [31] Priyabrata P., F. Banat; *J. Natural Gas Sci. Eng.*, 2014, 18, 227-236.
- [32] Lagergren S.; *Handlingar*, 1898, 24 (4) , pp. 1–39.
- [33] Zhuhong Ding a,c, Xin Hub,c, Yongshan Wand, Shengsen Wang c, Bin Gao; *Journal of Industrial and Engineering Chemistry*, 2016, 33, 239–245.
- [34] Ho Y.S., D.A. John Wase, C.F. Forster; *Water Res.*,1995, 29, 1327-1332.
- [35] Kun-Yi Andrew Lin, Yu-Ting Liu, Shen-Yi Chen; *Journal of Colloid and Interface Science*, 2016, 461, 79–87.
- [36] Dilip Kumar Mondal a, Barun Kumar Nandi b, M.K. Purkait; *Journal of Environmental Chemical Engineering*, 2013, 1, 891-898.
- [37] Özacar M. and I.A. Sengil; *Process Biochem.*,2005, 40, 565–572.
- [38] Moaaz K. Seliem, Sridhar Komarneni, Mostafa R. Abu Khadra; *Microporous and Mesoporous Materials*, 2016, 224, 51-57.
- [39] Malihe F., M.Beheshti, H. Sabzyan; *J. Molecular Liquids*, 2015, 211, 1060–1073.
- [40] Jingjing W., Z. Lia; *J Hazard. Mater.*, 2015, 300, 18–28.
- [41] Yu, K., Gu, C., Boyd, S.A., *et al.*; *Environ. Sci. Technol.*, 2012, 46, 8969–8975.
- [42] Yanping Z., N. Gao, Q. Wang, X. We; *Colloids and Surfaces A: Physicochem. Eng. Aspects*, 2015, 468, 114–121.
- [43] Qiong L., J.Yu, F. Zhou, X. Jiang; *Colloids and Surfaces A: Physicochem. Eng. Aspects*, 2015, 482, 306–314.
- [44] Mehmet, U.; *Microporous and Mesoporous Materials*, 2009, 119, 276–283.
- [45] Sushanta, D., Maity, A., Pillay, K., *J. Environ. Chem. Engineering*, 2014, 2, 260–272.
- [46] Singh, B., Das, S. K.; *Colloids and Surf. B: Biointer.*, 2013, 107,97– 106.