

Accumulation of Hg, Pb, Cr, and Fe in muscle and head of four fish species: Diplodus annularis, Zosterisessor ophiocephalus, Liza aurata and Caranx rhonchus

Ilhem Ketata Khitouni¹, Nourhène Boudhrioua Mihoubi², Abderrahmen Bouain³ and Faouzi Ben Rebah^{4*} ¹UR Biodiversité et Ecosystèmes Aquatiques, FSS, Sfax, Tunisia E-mail : ilhem_ketata@yahoo.fr ²UR Ecophysiologie et Procédés Agroalimentaires, ISB-Sidi Thabet, Tunisia E-mail : nourhene.boudhrioua@gmail.com ³UR Biodiversité et Ecosystèmes Aquatiques, FSS, Sfax, Tunisia E-mail : abderrah.bouain@fss.rnu.tn ⁴Laboratoire de Biochimie et de Génie Enzymatique des Lipases, ENIS, Sfax-Tunisia E-mail : benrebahf@yahoo.fr

ABSTRACT

Because heavy metal pollution was recently reported in the Gulf of Gabes (Tunisia), the concentration of selected elements (Hg, Pb, Cr and Fe) in the muscles and heads for fish species, *Diplodus annularis, Zosterisessor ophiocephalus, Liza aurata* and *Caranx rhonchus*, were determined using atomic absorption spectrometry. The element contents were identified to have the following decreasing sequence: Fe > Pb, Cr > Hg in both muscles and heads. However, levels of these elements were higher in heads but in trace in muscles. This indicates that, heads were contaminated but muscles were safe for the species caught from Gabes gulf area.

Keywords

Heavy metals, bioaccumulation, fish muscle, head



Council for Innovative Research

Peer Review Research Publishing System

Journal: JOURNAL OF ADVANCES IN BIOLOGY

Vol 5, No.1

editor@cirjab.com

www.cirjab.com, www.cirworld.com

559 | Page

June 20, 2014



INTRODUCTION

The fishery sector plays an important socioeconomic role in coastal countries. The world production of fishes and invertebrates is approximately 100 million tons annually. The average human consumption of fish is about 12 kg/person annually. In Tunisia, the fish production was estimated to about 100.451 tons and the coastal catch was about 22.662 tons [1]. This high consumption of seafood products is due to their significant amounts of different beneficial nutrients such as nutritional and digestible proteins, lipid-soluble vitamins, essential minerals and highly unsaturated fatty acids [2,3]. Most of these constituents have been shown to play important roles in human diet. As the fish stock tends to decrease, this natural food reserve should be preserved from contamination by toxic products as well as industrial waste. The effects of anthropogenic pollution on seawater become increasingly serious. One of the major pollution problems that pose serious health risk and environmental concern is the presence of heavy metals because of their toxicity, long persistence, bioaccumulation and biomagnifications in the food chain [4,5]. Fish samples can be considered as one of the most significant indicators in seawater systems for the estimation of metal pollution level [6,7,8]. In recent years, much attention has been directed to the concentrations of some inorganic elements in seawater fish [9,10,11]. The non-essential metals such as Hg, Cr and Fe have no known physiological function in fish, but can also be acquired from the diet. Both types of metals can originate from a variety of sources (domestic, agricultural and industrial sources) [12,13,14].

Generally, marine organisms absorb minerals from their diet and the surrounding water and deposit them in their skeletal tissues and organs. Various elements are widely known to be present in enzyme active centers that are responsible for the development of important functions in all animals; thus, marine-derived foods can serve as a good source of essential elements [15,16]. Additionally, fish and shellfish are good bioindicators of trace element contamination in the marine environment because they occupy different trophic levels and can exhibit large bioaccumulation factors. By studying fish and shellfish, the harmful effects of certain metals and metalloids to marine environment and to human health have been recognized [17,18]. For both the essential and toxic groups, elements corresponding to the transition and electronegative groups of the periodic table have been reported to be strongly bound to other constituents, whereas elements corresponding to electropositive groups, generally, remain dissolved in the ionic state in the cell medium [19]. Preliminary studies have indicated that the concentration of essential and toxic minerals in fish is influenced by a number of factors such as seasonal and biological differences (species, size, muscle, age, sex and sexual maturity), food source and environment (water chemistry, salinity, temperature and contaminants) [16,18].

The commercial and edible species have been widely investigated in order to check for those hazardous to protect human health. Hence, the aim of this study was to determine the seasonal variations of Hg, Pb, Cr and Fe in muscle and head of *Diplodus annularis, Zosterissor ophiocephalus, Liza aurata and Caranx rhonchus* species caught from Gabes gulf area (Tunisia).

MATERIALS AND METHODS

Fish material

Diplodus annularis, Zosterisessor ophiocephalus, Liza aurata and Caranx rhonchus species were caught from Gabes gulf area (Tunisia). Fishes were rapidly transported on ice to the laboratory for preparation to elemental analysis. The total length (12-14; 14-17; 20-30 and 16-21 cm, respectively) and weight (30-50, 30-60, 80-200 and 50-100 g, respectively) were mesured. Weight of samples was determined by using a precision scale 10^{-4} g (Sauter). Heads and muscles were dissected.

Mineral analysis

Each sample was dried at 105° C for 48 h to constant weight. 1 g of dried tissues were weighed and digested with concentrated HNO₃ at 120° C. When fumes were white and the solution was completely clear, the samples were cooled to room temperature and the tubes were filled to 5 ml with ultra pure water [20]. Sea water samples were stabilized at pH 2 with 1 mol/l HNO₃ prior to direct determination of total metal concentrations .The filtrates were analyzed by atomic absorption spectrophotometry (Perkin Elmer AAnalyst 200) [21].

Statistical analysis

Statistical analysis were performed by using SPSS software® version 11.0 (Statistical Package for Social Sciences). Values are expressed as mean \pm error deviation. Variance analysis was performed for Hg, Pb, Cr and Fe concentrations in both muscle and the head according to the factors sex, size of the fish species to determine the pertinent factors increasing the toxic element in muscle or head of fish (p < 0.05). Correlation matrixes were established between the measured variables (Hg, Pb, Cr and Fe). Honestly, significant difference (HSD) with ANOVA one factor was performed. Every factor presenting a p-value (p) inferior to 0.05 was considered significant.

RESULTS

Results of the seasonal variations of Hg, Pb, Cr and Fe in head and muscle of *D. annularis* were given in Table 1. The rough content orders of heavy metals in the head decreased in the sequence: Fe > Pb > Cr > Hg in head and in the sequence of Fe > Cr = Pb = Hg in the muscle. The Highest iron concentrations were found in the head of *D. annularis*, while the lowest levels were found in fish muscle. Fe was found to be the most abundant element in muscle. The higher values were obtained in autumn (3.28 and 3.19 mg / 1kg of fresh fish muscle for males and females respectively). Whereas, Hg, Cr and Pb were present at very low levels (inferior to 0.2 mg / kg of fresh fish muscle). Besides, the head of



D. annularis contained high levels of iron, the maximum values varied between 664.96 mg / kg (in spring) and 1815.93 mg / kg (in summer) for males; and between 843.04 mg / kg (in summer) and 1531.31 mg / kg (in autumn) for females. The maximum lead concentrations in the head were obtained in spring (286.34 mg / kg for males and 160.30 mg / kg for females). The head contained a higher level of Cr in summer (1815.93 mg / kg) for males and in autumn for females (1531.31 mg / kg). For Hg, values varied from 2.29 and 0.89 mg / kg (in summer) to 2.56 and 1.22 mg / kg (in winter) for male and female heads, respectively.

Table 1. Seasonal variation of Hg, Pb, Cr and Fe in head and muscle of D. annu	laris (ma / Ka of fresh fish)
Table 1. Seasonal valiation of ny, Fb, Cr and Fe in nead and muscle of <i>D. annu</i>	ians (ing / ky or nesh hsh).

		Winter	Spring	Summer	Autumn
Diplodus ann	<i>ularis</i> head	b		1	
Hg	Male	2.56±0.12a	2.77±0.20a	2.29±0.20a	2.50±0.08a
ТŊ	Female	1.22±0.10a	1.44±0.12a	0.83±0.16a	1.14±0.05a
Pb	Male	198.67±31.98a	286.34±24.89a	126.79±23.24a	54.67±8.82a
ΓIJ	Female	147.03±21.65a	160.30±12.94a	46.13±17.27a	72.33±8.27a
Cr	Male	73.90±2.72a	29.63±10.76a	114.73 ±10.11b	76.37±2.08a
CI	Female	35.29±1.02a	62.29±3.53a	33.43 ± 7.31ab	62.62±7.81b
Fe	Male	1712.17±207.23a	1664.96±251.22a	1815.93±163.38a	1728.15±51.20a
	Female	1076.36 ±144.13 a	1141.78±179.83a	843.04±127.59a	1531.31±108.89a
Diplodus ann	<i>ularis</i> mus	cle			
Hg	Male	0.02±0.10b	0.02±0.00b	0.03±0.00a	0.20±0.00b
ing	Female	0.02±0.00ab	0.01±0.00a	0.03±0.00b	0.19±0.00c
Pb	Male	0.17± 0.00d	0.11±0.00c	0.11±0.00b	0.10 ± 0.00a
T D	Female	0.16±0.00c	0.11±0.00b	0.11±0.00b	0.04 ± 0.00a
Cr	Male	0.20±0.00b	0.07±0.00a	0.20±0.00b	0.20±0.00b
OI IIII	Female	0.20±0.00b	0.09±0.00a	0.20±0.00b	0.20±0.00b
Fe	Male	1.12±0.01b	0.92±0.00a	1.17±0.00c	3.28±0.00d
ге	Female	1.10±0.07a	110±0.02a	1.07±0.04a	3.19±0.04b

In the case of, *Z. ophiocephalus,* head and muscle presented the same rough content orders (in comparison to *D.annularis* (Table 2). In heads, the Hg element varied between 1.63 (in winter) and 2.20 mg / kg (in autumn) for males and from 0.76 (in summer) to 1.03 mg / kg (in winter) for females. The Cr values were higher in autumn (130.79 and 41.12 mg / 100 g for males and females head, respectively). For lead, the concentrations were high in winter (228.04 mg / kg for males and 104.24 mg / kg for females). Moreover, *Z. ophiocephalus* heads presented high iron concentration with values ranged between 1724.33 (in winter) and 2352.96 mg / kg (in summer) for males and between 653.32 (in summer) and 1393.12 mg / kg (in autumn) for females. In addition to that, the muscle of *Z. ophiocephalus* showed lower values of Hg, Cr and Pb (inferior to 0.2 mg/ kg). Fe was found to be the most abundant element in muscle with maximum values in winter (4.80 and 4.60 mg/ kg for males and females, respectively).

Table 2. Seasonal variation of Hg. Pb. Cr and Fe in head and muscle of Z. ophiocephalus (mg / Kg of fresh fish)

		Winter	Spring	Summer	Autumn
Z. ophiocep	<i>bhalus</i> head				
Hg	Male	1.63±0.36a	2.15±0.28b	2.12±0.38ab	2.20 ±0.31ab
1.19	Female	1.03±0.15ab	0.91±0.20b	0.76 ±0.20a	1.00 ±0.06a
Pb	Male	228.04±46.17ab	237.75 ±40.96b	118.74 ±24.85ab	117.80± 3.77a
	Female	104.24±24.85ab	7.990 ±14.39b	49.53±5.08a	79.26 ±10.70a
Cr	Male	63.24±12.97a	59.56±0.32ab	93.38± 20.91c	130.79 ±18.19bc
	Female	33.36±01.59a	20.33±5.86a	34.02 ±10.69a	41.12±1.92a
Fe	Male	1724.33±375.57a	1793.32±171.54ab	2352.96±126.47b	1994.58±407.10ab



	Female	814.89±130.06a	718.34±162.49a	65.332±10.486a	139.312±23.71a			
Z. ophiocepha	Z. ophiocephalus muscle							
Hg	Male	0.20 ± 0.00c	0.02±00.00a	0.05±0.00b	0.20±0.00c			
iig	Female	0.20 ± 0.00c	0.03±00.00a	0.04±0.00b	0.30±0.00d			
Pb	Male	0.22 ± 0.00d	0.12±00.00b	0.17±0.00c	0.07±0.00a			
	Female	0.08 ± 0.00a	0.22±00.01c	0.13±0.00b	0.15±0.00b			
Cr	Male	0.20 ± 0.00c	0.06±00.00a	0.10±0.01b	0.20±0.00c			
	Female	$0.20 \pm 0.00b$	0.10±00.00a	0.10±0.00a	0.30±0.00c			
Fe	Male	4.80 ± 0.01d	0.64±00.04a	2.17±0.03b	2.42±0.01c			
	Female	4.60 ± 0.20d	1.40±00.09b	1.02±0.06a	3.84±0.04c			

Similarly to *D.annularis* and *Z.ophiocephalus*, the seasonal variation of Hg, Pb, Cr and Fe in head and muscle of *L. aurata showed* the same rough content orders (Table 3). For *L. aurata* heads, Hg varied between 1.11 (in winter) and 1.97 mg / kg (in spring) for males and from 0.78 (in autumn) to 0.98 mg / kg (in summer) for females, respectively. Also, Cr varied between 32.18 (in winter) and 71.17 mg / kg (in summer) for males and from 21.91 (in spring) to 58.05 mg / kg (in autumn) for females. However, the maximum lead concentration was observed in spring (17.674 and 8.443 mg / kg for males and females, respectively. In addition, the highest iron values were showed in summer (652.23 and 923.85 mg / kg for males and females, respectively). In muscle, Hg, Cr and Pb were low (inferior to 0.2 mg/ kg). Only in winter, the lead presented 0.49 mg /kg in muscle; however Fe was found to be the most abundant element with maximum values in winter (2.07 and 1.99 mg / kg for males and females, respectively).

		Winter	Spring	Summer	Autumn
L. aurata he	ad		0		
Hg	Male	1.11±0.31a	1.97±0.15a	1.66±0.25a	1.74±0.25a
ng	Female	0.89±0.20 a	0.94±0.24a	0.98±0.21a	0.78±0.22a
Pb	Male	113.28±28.75a	176.74±16.72a	83.52±7.64a	52.99±6.70a
T D	Female	63.76±16.50a	84.43±15.19a	49.51±17.05a	66.51±16.09a
Cr	Male	32.18±7.66a	50.65±7.38a	71.17±5.56a	55.59±3.38b
CI	Female	33.90±4.32ab	21.91±7.54a	35.24±7.15ab	58.05±7.56b
Fe	Male	304.95±81.57a	569.87±120.56a	652.23±39.75a	406.95±73.30a
re	Female	531.39±80.81a	406.14±0.67a	923.85±103.51a	882.70±111.47b
<i>L. aurata</i> mu	iscle				
Ha	Male	0.07±0.00b	0.01±0.00a	0.01±0.00a	0.13±0.00c
Hg	Female	0.10±0.00c	0.05 10 ⁻¹ ±0.00a	0.01±0.00b	0.10±0.00c
Pb	Male	0.03±0.00a	0.07±0.00d	0.05±0.00b	0.05±0.00c
FU	Female	0.49±0.00c	0.03±0.00a	0.04±0.00b	0.04±0.00b
Cr	Male	0.10±0.00c	0.03±0.00a	0.09±0.00b	0.10±0.00c
G	Female	0.10±0.00b	0.02±0.00a	0.10±0.00b	0.10±0.00b
Fe	Male	2.07±0.01d	0.92±0.02b	0.62±0.02a	1.91±0.00c
ге	Female	1.99±0.00d	0.30±0.00a	0.55±0.02b	1.74 ±0.02c

Table 3. Seasonal variation of Hg. Pb. Cr and Fe in head and muscle of L. aurata (mg / Kg of fresh fish).

Results related to *C. rhonchus* are presented in Table 4. Generally, both head and muscle of *C. rhonchus* presented the same rough content orders (Fe > Pb >Cr> Hg). In heads, the Hg element varied between 1.18 (in autumn) and 2.41 mg / kg (in spring) for males and from 0.65 (in spring) to 0.87 mg / kg (in autumn) for females. For Cr (in heads), values varied between 47.71 (in winter) and 105.15 mg / kg (in summer) for males and from 25.47 (in spring) to 39.44 mg / kg (in autumn) for females. For lead (in heads), values varied between 95.94% (in summer) and 286.04 (in spring) for males and from 40.35 mg / kg (in summer) to 113.87 (in winter) for females. Maximum iron values were presented in winter (1027.61)



and 861.03 mg / kg for males and females head, respectively). For *C. rhonchus* muscle, Hg, Cr and Pb elements were low (inferior to 0.3 mg/ 100 g). Fe was found to be the most abundant in muscles with maximum values obtained in winter (3.10 and 2.69 mg / kg for males and females, respectively). The elements levels were high in head compared with muscles.

Generally, Hg values in head of all fish species do not exceed 2.77 mg / kg. This value is recorded in spring for *D. annularis* males (Hg values ranged between 0.83 and 2.77 mg / kg). Cr reached 130.79 mg / kg in the case of *Z. ophiocephalus* (in autumn). However, Pb is moderately high for the four species with values ranging between 46.13 (in summer) and 286.34 mg / kg (in spring) for *D. annularis* females. The Fe contents were high and varied between 304.95 (*L. aurata* males in winter) to 2352.96 mg / kg (*Z. ophiocephalus* males in summer). The variation of Hg, Pb, Cr in muscle of all fish species was low (inferior to 0.3 mg/ 100 g). But, the Fe level varies between 0.30 mg / kg for *L. aurata* femelles (in spring) and 4.80 mg / kg for *Z.ophiocephalus* males.

		Winter	Spring	Summer	Autumn
C. rhonch	hus head				
Hg	Male	1.77±0.36a	2.41±0.11a	1.44±0.18a	1.18±0.31a
	Female	0.76±0.21 a	0.65±0.05a	0.84±0.20a	0.87±0.22a
Pb	Male	192.99±33.98 a	286.04±1.95a	95.94±23.56a	120.24±34.19a
	Female	113.87±26.85 a	57.45±8.92a	40.35±5.40a	60.05±22.44 a
Cr	Male	47.71±2.97ab	52.75±0.75a	105.15±17.85ab	71.33±1.46b
	Female	28.29±11.17 a	25.47±6.76a	38.42±15.31a	39.44±4.26a
Fe	Male	1027.61±204.87a	853.75±146.24a	738.96±68.16 a	738.33±132.34 a
	Female	861.03±114.06a	740.45±123.21a	1012.37±113.47 a	634.76±59.95 a
C. rhonch	hus muscle				
Hg	Male	0.10 ±0.01b	0.00a	0.01±0.00a	0.10±0.00b
	Female	0.10 ±0.00b	0.00a	0.01±0.00a	0.10±0.00b
Pb	Male	0.01±0.00a	0.04± 0.00b	0.05±0.00c	0.05±0.00c
	Female	0.04±0.00b	0.05± 0.00c	0.05±0.00c	0.00a
Cr	Male	0.30±0.00c	0.02± 0.00a	0.10±0.00b	0.10±0.00b
	Female	0.20±0.00c	0.04± 0.00a	0.10±0.00b	0.20±0.00c
Fe	Male	3.10±0.00d	0.44± 0.03a	0.63±0.03b	1.62±0.01c
	Female	2.69±0.04c	0.52± 0.01a	0.62±0.02a	2.22±0.05b

As indicated in table 5, the variation of heavy metals (Hg, Pb, Cr and Fe) in muscle was significant as a function of species, season, sexes and the interactions species season, species season, sexes and species x season x Sex ($p<10^{-3}$). Only the sex factors, it is not significant in iron variation (P<0.05).

Table 5. Effect of sex and species of fish and season factors on the variation of heavy metals contained in the
heads and muscles of of D. annularis. Z. ophiocephalus. L. aurata and C. rhonchus

		Head		Muscle	
Factors	Content	F (Fisher number)	p (p-value)	F (Fisher number)	p (p-value)
Species	Hg	52.75	<10 ⁻³	1734.05	<10 ⁻³
	Pb	15.99	<10 ⁻³	1480.39	<10 ⁻³
	Cr	56.71	<10 ⁻³	1581.26	<10 ⁻³
	Fe	95.78	0.03	2243.15	<10 ⁻³
	Hg	2.31	0.08	5395.86	<10 ⁻³
Season	Pb	16.79	<10 ⁻³	886.78	<10 ⁻³
	Cr	20.82	<10 ⁻³	3680.27	<10 ⁻³



					4 9 - 3
	Fe	2.48	0.07	6208.83	<10 ⁻³
	Hg	4.67	0.03	361.78	<10 ⁻³
Sex	Pb	1.13	0.29	235.60	<10 ⁻³
JEX	Cr	0.57	0.45	126.06	<10 ⁻³
	Fe	4.42	0.03	0.84	0.36
	Hg	1.13	0.35	394.19	<10 ⁻³
Species × Season	Pb	2.23	0.03	472.62	<10 ⁻³
Species x Season	Cr	3.29	<10 ⁻³	438.89	<10 ⁻³
	Fe	3.38	<10 ⁻³	920.15	<10 ⁻³
	Hg	1.85	0.14	809.54	<10 ⁻³
Species × Sex	Pb	2.90	0.04	498.75	<10 ⁻³
Opecies x Dex	Cr	3.20	0.03	63.81	<10 ⁻³
	Fe	5.68	<10 ⁻³	56.35	<10 ⁻³
	Hg	0.06	0.96	238.91	<10 ⁻³
$\mathbf{Sex} \times \mathbf{Season}$	Pb	0.87	0.46	344.92	<10 ⁻³
	Cr	0.47	0.70	243.31	<10 ⁻³
	Fe	0.34	0.79	183.97	<10 ⁻³
Species × Season × Sex	Hg	1.27	0.27	257.83	<10 ⁻³
	Pb	0.74	0.67	881.94	<10 ⁻³
	Cr	3.14	<10 ⁻³	109.12	<10 ⁻³
	Fe	1.33	0.23	140.08	<10 ⁻³

In heads, Pb and Cr varied significantly as a function of species, season and the interaction species×season and species×season action of Cr was significant with the interaction Species×Season×Sex (P< 0.05). However, Hg variation is significant as a function of species and sex factors (P< 0.05). The factors species, sex and the interaction species×season and species×season and species×season and species×season and species×season and species×season action of Fe (P< 0.05). The factors species sex have significant effect on the variation of Fe (P< 0.05). The factors species sex have significant effect on the variation of Fe (P< 0.05).

DISCUSSION

In the present work only fish muscles and heads were evaluated for the elemental concentrations. Iron, lead, mercury and chrome were selected from the viewpoint of the industry type near the golf of Gabes and the metal pollution anticipated. Although many researchers have presented the elemental contents in various tissues, such as liver, kidneys, gills, gonads and muscles of fish [11,22]. Among the analyzed elements, Fe was found to be the most abundant, whereas Hg, Cr and Pb were presented at very low levels in muscles. Fe, Hg, Pb and Cr found in the fish muscles are at acceptable levels (below the threshold) [23]. These results are consistent with those of other studies [24,25] concerning fish muscles from the lagoon Tuzla. These authors reported significant seasonal variations.

Higher heavy metal values were observed in heads compared with muscles. The elemental head values were superior to the threshold [23]. The ranges of international standards for fish are (in mg/ 100 g): 0.0055 ± 0.0006 for Hg; 0.003 ± 0.0015 for Pb; 0.018 ± 0.0055 % for Cr and 3.65 ± 0.35 for Fe. The rough content orders were: iron > lead>chromium> mercury. These sequences were the same as those obtained in Malibu Lagoon, California and Dhanmondi Lake in Bangladesh [26,27]. Data related Fe, Pb, Cr and Hg contents found in the present study are in agreement with mean values reported for muscles of most fish species [16,17,19]. It is well known that muscle is not an active tissue in accumulating heavy metals [28]. The greater presence of heavy metals in the gills than in muscle is probably due to the direct exposure of this organ to the water and thus to toxic compounds [29]. The least abundant metals in muscle were Hg, Cr and Pb. These values are similar to those reported by [30] in four fish species caught adjacent to Raine Island in northern Britain. Iron is quantitatively the most important in the muscle regardless of fish species and season. It is involved in the base metabolism [31]. Iron contents have exposed fluctuations depending on the species, the sex of fish and season. Fe serves as a carrier of oxygen to the muscle from the lungs by red blood cell haemoglobin, as a transport medium for electrons within cells and as an integrated part of important enzyme systems in different tissues [32,33]. The average intake of Fe is too low, although many people receive more than 18 mg per day, which is the recommended dietary allowance (RDA) [34]. However, lead showed no physiological role in fish [35], but it can be absorbed by the gastrointestinal tract [36,37,38]. It showed high values in head for all species. Chromium is essential for normal carbohydrate and lipid metabolism. The role of chromium in glucose metabolism has been reported for poults and



mammals. It is considered to be a cofactor for insulin activity and part of an organic tolerance factor [39,40]. Also, it was reported the influence of dietary chromium on glucose metabolism of fish [41]. Hence, chromium salts improved glucose utilization and inhibited gluconeogenesis, probably by modulating the endogenous insulin activity. Supplemental dietary chromium increased the weight gain, energy deposition and liver glycogen content in tilapia fed glucose diets [42]. The biological availability of this element depends on its characteristic ability to form coordination compounds and chelates. It occurs in food as part of a biologically active molecule and as inorganic trivalent chromium. The contamination by heavy metals is in direct relation with the diet. *D. annularis* feeds on worms, crustaceans, molluscs, echinoderms, hydrozoans and algae that absorb high levels of heavy metals. *L. aurata* feeds on small plants, invertebrates and various detritus dig into the bottom sediments and filter through their gill rakers. However, *Z. ophiocephalus* feeds on small fish and crustaceans [43,44]. It is well-known that a number of factors such as sex, age, season, spawning period, variability of food habitats, pollutant exposure and phylogenetical differences in regulatory mechanisms, may influence the uptake, retention and bioaccumulation of trace contaminants in fish tissues. In the case of *Z. ophiocephalus*, a near-bottom feeder, sediments may also play an important role as a source of contaminated food [45]. *C. rhonchus* feeds on small fish and invertebrates [43,44]. In comparison with other species, *C. rhonchus* is less contaminated by heavy metals and their diet is responsible for the lower contamination.

CONCLUSION

The study of the seasonal variation of the heavy metals, from the Gabes Gulf (Tunisia) is important and useful to study the effect of pollution on the consumable fish. The significant seasonal differences observed in the mineral elements are mainly related to diet and environmental factors. Based on the present study, it is recommended to do not eat fish heads.

REFERENCES

- [1] DGPA., 2009. Annuaire des statistiques des produits de la pêche. Direction général de la pêche et de l'aquaculture. Ministère de l'agriculture Tunisie, 114p.
- [2] Aitken, A., Mackie, I., Merrit, J., and Windsor, M. 1982. Fish: Handling and Processing. Ministry of Agriculture, Fisheries and Food, Torry Research Station, Edinburgh, Scotland, UK, pp. 2–19.
- [3] Simopoulos, A. 1997. Nutritional aspects of fish. In: Luten, J., Börrensen, T., Oehlenschlager, J. (Eds.), Seafood from Producer to Consumer, Integrated Approach to Quality. Elsevier Science, London, UK, pp. 589–607.
- [4] Eisler, R., 1988. Zink Hazards to fish, Wildlife and Invertebrates: a synoptic review. US Fish Wildlife Serv. Biology of Reproduction, 85.
- [5] Hei, A., and Sarojnalini, C.H. 2012. Study Of Protein Quality Of Some Fresh And Smoke-Dried Hill Stream Fishes From Manipur, India. New York Science Journal. 2012;5(11).
- [6] Barak, N.A., and Mason, C.F., 1990. Mercury, cadmium and lead in eels and roach: the effects of size, season and locality on metal concentrations in flesh and liver. Science of the Total Environment 92, 249–256.
- [7] Evans, D.W., Dodoo, D.K., and Hanson, P.J. 1993. Trace element concentrations in fish livers: implications of variations with fish size in pollution monitoring. Marine Pollution Bulletin, 26, 329–34.
- [8] Rashed, M.N. 2001. Monitoring of environmental heavy metals in fish from Nasser lake. Environment International, 27: 27–33.
- [9] Farkas, A., Sala´nki, J., and Speczia, R. A. 2003. Age and size-specific patterns of heavy metals in the organs of freshwater fish Abramis brama L. populating a low-contaminated site. Water Research, 37, 959–964.
- [10] Mansour, S. A., and Sidky, M. M. 2002. Ecotoxocological studies. 3. Heavy metals contaminating water and fish from Fayoum Governorate, Egyptian Food Chemistry, 78, 15–22.
- [11] Moiseenko, T.I., and Kudryavtseva, L. P. 2001. Trace metal accumulation and fish pathologies in areas affected by mining and metallurgical enterprises in the Kola Region, Russia. Environmental Pollution, 114, 285–297.
- [12] World Health and Organisation. 1995. Environmental Health Criteria. International Programme on Chemical Safety. World Health Organisation, Geneva, 165p.
- [13] Purcell, T.W., and Peters, J.J. 1998. Sources of silver in the environment. Environmental Toxicology and Chemistry 17(4), 539-546.
- [14] Wood, C.M. 2001. Toxic responses of the gill. In: Schlenk, D.W., Benson,W.H. (Eds.), Target Organ Toxicity in Marine and Freshwater Teleosts, Organs, vol. 1. Taylor and Francis, Washington, DC, USA, pp. 1–89.
- [15] Johnson, M., and Fischer, J., 1994. Role of minerals in protection against free radicals. Food Technology 48 (5), 112– 120.
- [16] Lal, S. 1995. Macro and trace elements in fish and shellfish. In: Ruiter, A. (Ed.), Fish and Fishery Products: Composition, Nutritive Properties and Stability. CAB International, Wallingford, CN, USA, pp. 187–214.
- [17] Engman, J., and Jorhem, L. 1998. Toxic and essential elements in fish from Nordic waters, with the results seen from the perspective of analytical quality assurance. Food Additives and Contaminants 15, 884–892.

- [18] Noël, L., Chafey, C., Testu, C., Pinte, J., Velge, P., and Guerin, T. 2011. Contamination levels of lead, cadmium and mercury in imported and domestic lobsters and large crab species consumed in France: differences between white and brown meat. Journal of Food Composition 24, 368–375.
- [19] Piclet, G., 1987. Le poisson aliment. Composition-interêt nutritionnel. Cahier de nutrition et diétetique 22, 317–336.
- [20] Warchalowska-S´liwa, E., Niklin´ska, M., Gorlich, A., Michailova, P., and Pyza, E., 2005. Heavy metal accumulation, heat shock protein expression and cytogenetic changes in *Tetrix tenuicornis* (L.) (Tetrigidae, Orthoptera) from polluted areas. Environmental Pollution 133, 373–381.
- [21] Bervoets, L., and Blust, R. 2003. Metal concentrations in water, sediment and gudgeon (Gobio gobio) from a pollution gradient: relationship with fish condition factor. Environmental Pollution126, 9-19.
- [22] Mzimela, H.M., Wepener, V., and Cyrus, D.P. 2003. Seasonal variation of selected metals in sediments, water and tissues of the groovy mullet, *Liza dumerelii* (Muglidae) from the Mhlathuze Estuary, South Africa. Marine Pollution Bulletin 46: 659-676.
- [23] IAEA-407. 2003. Trace éléments and methylmercury in fish tissue. International Atomic Energy Agency. Analytical Quality Control Services. Wagramer Strasse 5, P.O. Box 100, A-1400 Vienna, Austria, 4p.
- [24] Miller, P.A., Munkittrick, K.R., and Dixon, D.G. 1992. Relationship between concentrations of copper and zinc in water, sediment, benthic invertebrates and tissues of white sucker (*Catastomus commersoni*) at metal-contaminated sites. Canadian Journal of Fisheries and Aquatic Sciences 49, 978–985.
- [25] Dural, M., Goksu, M.Z.L., and Ozak, A.A. 2007. Investigation of heavy metal levels in economically important fish species captured from the Tuzla lagoon. Analytical, Nutritional and Clinical Methods. Food Chemistry 102, 415–421.
- [26] Moeller, A., MacNeil, S.D., Ambrose, R.F., and Hee, S.S.Q. 2003. Elements in fish of Malibu Creek and Malibu Lagoon near Los Angeles, California. Marine Pollution Bulletin 46, 424–429.
- [27] Begum, A., Amin, M.N., Kaneco, S., and Ohta, K. 2005. Selected elemental composition of the muscle tissue of three species of fish, *Tilapia nilotica*, *Cirrhina mrigala* and *Clarius batrachus*, from the fresh water Dhanmondi Lake in Bangladesh. Food Chemistry 93, 439–443.
- [28] Sunlu, U., Ozdemir, E., and Basaran, A. 2001. The red mullet *Mullus barbatus* (Linnaeus 1758) as an indicator for heavy metal pollution in Izmir Bay (Turkey). in: 36th Ciesm Congress Proceedings, Monte Carlo, Monaco.
- [29] De Souza, S.N., and Naqvi, S.W.A. 1979. Metal concentrations in the grew mullet from (*Mugil cephalus*) Visakha patnam. National Institute of Oceanography, Dona Paula, India 12 (4), 259–262.
- [30] Rayment, G.E., and Barry, G.A. 2000. Indicator tissues for heavy metal monitoring additional attributes. Marine Pollution Bulletin 41(7–12), 353–358.
- [31] Mnari Bhouri, A., Bouhlel, I., Chouba, L., Hammami, M., El Cafsi, M., and Chaouch, A. 2010. Total lipid content, fatty acid and mineral compositions of muscles and liver in wild and farmed sea bass (*Dicentrarchus labrax*). African Journal of Food Science 4(8), 522 530.
- [32] Wagner, A., and Boman, J., 2003. Biomonitoring of trace elements in muscle and liver tissue of freshwater fish. Spectrochimica Acta. Part B. 58, 2215-2226.
- [33] Camara, F., Amaro, M.A., Barbera, R., and Clemente, G., 2005. Bioaccessibility of minerals in school meals: comparison between dialysis and solubility methods. Food Chemistry 92, 481-489.
- [34] Tolonen, M., 1990. Vitamina and minerals in health and nutrition, Ellis Horword, London, pp. 152-188.
- [35] Adeola, A., Ojo, L., and Wood, C.M., 2007. In vitro analysis of the bioavailability of six metals via the gastro-intestinal tract of the rainbow trout (*Oncorhynchus mykiss*). Aquatic Toxicology 83, 10–23.
- [36] Crespo, S., Nonnotte, G., Colin, D.A., Leray, C., Nonnotte, L., and Aubree, A., 1986. Morphological and functional alterations induced in trout intestine by dietary cadmium and lead. Journal of Fish Biology 28, 69–80.
- [37] Mount, D.R., Barth, A.K., Garrison, T.A., Barten, K.A., and Hockett, J.R. 1994. Dietary and waterborne exposure of rainbow trout (*Oncorhynchus mykiss*) to copper, cadmium, lead and zinc using a live diet. Environmental Toxicology and Chemistry 13, 2031–2041.
- [38] Alves, L., Glover, C.N., and Wood, C.M. 2006. Dietary Pb accumulation in juvenile freshwater rainbow trout (*Oncorhynchus mykiss*). Archives of Environmental Contamination and Toxicology 51, 615–625.
- [39] Anderson, R. 1981. Nutritional role of chromium. Science of the Total Environment 17, 13–28.
- [40] Steele, N.C., and Rosebrough, R.W. 1981. Effect of trivalent chromium on hepatic lipogenesis by the turkey poults. Poultry Science 60, 617-622.
- [41] Hertz, Y., Madar, Z., Hepper, B., and Gertler, A. 1989. Glucose metabolism in the common carp (*Cyprinus carpio* L.1): the effects of cobalt and chromium. Aquaculture 76, 255-267.



- [42] Shiau, S.Y., and Lin, S.F. 1993. Effect of supplemental dietary chromium and vanadium on the utilization of different carbohydrates in tilapia, *Oreochromis niloticus* X 0. *aureus*. Aquaculture 110, 321-330.
- [43] Fischer, W., Bauchot, M. L., and Schneider, M. 1987. *Fiches F.A.O.* d'identification des espèces pour les besoins de la pêche "Révision" Méditerranée et Mer noire. Zone de pêche 37. Volume I et II. Vertébrés. Rome, F.A.O ; 1530 p.
- [44] Bradaï, M.N. 2000. Diversité du peuplement icthyque et contribution à la connaissance des sparidés du golfe de Gabès ; Thèse es-sciences, Université de Sfax (Tunisie). 600 p.
- [45] Croteau, M.N., Luoma, S.N., and Stewart, A.R. 2005. Trophic transfer of metals along freshwater food web: evidence of cadmium biomagnifications in nature. Limnology and Oceanography 50 (5), 1511–1519.

